# Phenomics: Expanding the Role of Clinical Evaluation in Genomic Studies

Matthew B. Lanktree, BSc,\*† Reina G. Hassell, MSc,† Piya Lahiry, MSc,\*† and Robert A. Hegele, MD, FRCPC, FACP\*†

Abstract: With advances in high-throughput genotyping technologies, the rate-limiting step of large-scale genetic investigations has become the collection of sensitive and specific phenotype information in large samples of study participants. Clinicians play a pivotal role for successful genetic studies because sound clinical acumen can substantially increase study power by reducing measurement error and improving diagnostic precision for translational research. Phenomics is the systematic measurement and analysis of qualitative and quantitative traits, including clinical, biochemical, and imaging methods, for the refinement and characterization of a phenotype. Phenomics requires deep phenotyping, the collection of a wide breadth of phenotypes with fine resolution, and phenomic analysis, composed of constructing heat maps, cluster analysis, text mining, and pathway analysis. In this article, we review the components of phenomics and provide examples of their application to genomic studies, specifically for implicating novel disease processes, reducing sample heterogeneity, hypothesis generation, integration of multiple types of data, and as an extension of Mendelian randomization studies.

**Key Words:** phenomics, genetics, phenotype, cluster analysis, Mendelian randomization

(J Investig Med 2010;58: 700-706)

enome-wide association studies (GWASs), examining a wide range of discrete and quantitative phenotypes, have now been performed in many large cohorts. Mega meta-analyses, including samples of more than 75,000 subjects and often including more than 100 authors represent Herculean collaborative efforts to identify phenotype-associated genetic variants. Although the associated loci may be biologically valid and important, the frequency and effect size of the identified variants will become smaller because very large samples are required to confidently identify the phenotype-genotype asso-

From the \*Departments of Medicine and Biochemistry, Schulich School of Medicine & Dentistry; and †Robarts Research Institute, University of Western Ontario, London, Ontario, Canada.

Received December 10, 2009, and in revised form February 8, 2010.

Accepted for publication February 8, 2010.

Reprints: Robert A. Hegele, MD, FRCPC, FACP, Blackburn Cardiovascular Genetics Laboratory, Robarts Research Institute, University of Western Ontario, London, Ontario, Canada N6A 5K8. E-mail: hegele@robarts.ca.

Supported by the Canadian Institutes for Health Research (MOP-13430; MOP-79533), the Heart and Stroke Foundation of Ontario (NA-6059; T-6018), Genome Canada through Ontario Genomics Institute, and the Pfizer Jean Davignon Distinguished Cardiovascular and Metabolic Research Award.

M.B.L. is supported by the Canadian Institute of Health Research (CIHR) MD/PhD Studentship Award. P.L. is supported by the CIHR Scriver Family MD/PhD studentship award. Both M.B.L. and P.L. are supported by the University of Western Ontario MD/PhD Program and are CIHR Fellows in Vascular Research.

The authors have no conflicts of interest to declare.

Copyright  $\ensuremath{\mathbb{C}}$  2010 by The American Federation for Medical Research

ISŜN: 1081-5589

DOI: 10.231/JIM.0b013e3181d844f7

ciation signal. Thus, precision in subject ascertainment and phenotype definition should be key elements in the investigation of the genetic basis of complex diseases.<sup>2–4</sup>

What are the main obstacles to successful identification of genetic contributors to disease? Major culprits for reducing the signal-to-noise ratio and obscuring phenotype-genotype associations are measurement error and study heterogeneity.5 Geneticists are now accustomed to technologies with extraordinarily low imprecision: genotype miscall rates are typically less than 0.5%. However, measurement error in phenotypes has received substantially less attention. Confounding factors in genetic studies, such as pleiotropy, incomplete penetrance, epistasis, and allelic and locus heterogeneity, are unavoidable and need to be considered in study design.<sup>5</sup> However, study heterogeneity originating from clinical sources, such as phenocopy or multiple pathways leading to a common disease phenotype, can potentially be mitigated. Careful measurement not only of the major phenotype of interest but also of environmental exposures and subphenotypes, also known as biomarkers, endophenotypes, subclinical traits, or attributes, can provide valuable clues to the pathophysiology of each individual case.

Phenomics is the systematic measurement and analysis of qualitative and quantitative traits, including clinical, biochemical, and imaging methods, for the refinement and characterization of a phenotype. The importance of accurate phenotype determination in association studies has already been proven theoretically, and yet, phenomics has not generally received the same scientific and technological attention as genomic analysis. As attention shifts from increasing sample size to increasing diagnostic precision in the postgenomic era, the clinician, in particular, can play a pivotal role for the collection of accurate and complete phenotype information to ensure that genetic investigations have maximal power to identify consistent genotype-phenotype associations. Here, we further define and describe applications of "phenomics" and provide examples demonstrating its utility.

#### **PHENOMICS**

In its simplest form, a retrospective genetic association study design involves the comparison of allele frequencies between a collection of affected cases and unaffected controls.<sup>3</sup> Because additional covariates can increase the likelihood of an individual becoming a case, attempts to match the cases and controls for covariates such as age and sex are common. By extending and increasing the sophistication of techniques for describing and quantifying the phenotype of cases and controls, phenomics holds the promise of similarly helping to reduce study heterogeneity. Current evidence suggests that common complex disorders are in truth extremes of quantitative traits, in which additional information can be gained through precise, quantitative phenotypic description.<sup>10</sup> Phenomics is composed of 2 separate components: "deep phenotyping" referring to a strategic and comprehensive approach toward data acquisition

**TABLE 1.** Deep Phenotyping for a Comprehensive Phenomic Assessment

Components	<b>Example Methods</b>	Example Sample Data
Medical history	Physician history, questionnaires	Age, age of onset, diet, allergies, risk factors, triggering factors, environmental exposures, complete review of systems, family history
Physical examination	Anthropometric measurement, medical devices	Height, weight, hip-to-waist circumference, BMI, BP, pulse, heart sounds, retinal appearance, acanthosis nigricans
Quantitative clinical investigations	Biochemical, serological, immunological, pathological	CBC, insulin, fasting glucose, $Hb_{A1c}$ , lipid profile, leptin, adiponectin, CRP, ESR, rheumatoid factor, IL-2, blood clotting factors, ferritin, liver enzymes
Imaging	Ultrasound, x-ray, CT, MRI, PET, angiogram	Sc-fat mass, liver fat content, cardiac hypertrophy, carotid plaque area, intima media thickness, functional or perfusion scans
Sampling	Phlebotomy, biopsies, surgical specimens	Plasma, lymphocytes, tissue biopsies, resected tissue

BMI indicates body mass index; BP, blood pressure; CBC, complete blood count; CRP, C-reactive protein; CT, computed tomography scan; ESR, erythrocyte sedimentation rate; Hb<sub>A1c</sub>, glycosylated hemoglobin; IL-2, interleukin-2; MRI, magnetic resonance imaging; PET, positron emission tomography; sc, subcutaneous.

and "phenomic analysis" referring to the evaluation of patterns and relationships between individuals with related phenotypes and between genotype-phenotype associations.<sup>2,11</sup>

#### **DEEP PHENOTYPING**

Deep phenotyping involves the development of a complete picture of each study participant through the strategic collection of a broad range of high-resolution phenotypes (Table 1). Phenotype resolution, or granularity, is the level of detail afforded to phenotypic definition. 12 The goal of deep phenotyping is to characterize further as many of the contributing factors to the "case" definition as possible, which may allow for the removal or correction of heterogeneity among research subjects. In general, continuous quantitative phenotypes can better differentiate between marginal and severe cases and generally allow for more powerful statistical comparisons than do qualitative traits. Deep phenotyping begins with the collection of a thorough medical history, including a detailed account of environmental exposures, a complete review of systems, and collection of family history. Measurement of disease progression, an extensive panel of risk factors, and alternative measures of shared disease pathways should be used to provide the most detailed phenotype possible. By extending the detail, accuracy, and context of trait acquisition, and then linking components, a more complete phenotype can be generated for each individual.<sup>11</sup>

In their day-to-day work focusing on diagnosis, clinicians use technologies that probe intermediate markers in pathways underlying illness, including biochemical, serological, histopathological, and noninvasive imaging methods. These assays

can be static, such as biochemical analysis of plasma or tissue samples, or dynamic, such as provocative tests followed by serial sampling performed in clinical investigation units. For instance, in cardiovascular research, static biochemical phenotypes include serum concentrations of insulin, fasting glucose, triglycerides, low-density lipoprotein (LDL) and high-density lipoprotein cholesterol levels. Dynamic phenotypes include challenge by a stimulus or provocation of a response, such as postprandial excursion of plasma metabolites or measurement of insulin sensitivity using a euglycemic insulin clamp. Using serial measurements to monitor progression or regression of the phenotype, either in response to treatment or simply over time, becomes a further dimension of the phenotype. Thus, the same principles clinicians use to develop specific diagnoses can be applied to phenomic research applications.

In addition to data quality and granularity, deep phenotyping requires optimal data-gathering conditions to reduce technical variability and thus maximize the chance that the observed variation is biological in origin. Whenever possible, quantitative phenotyping methods should be performed using standard operating procedures and be validated against reference standards, with clear performance metrics, such as measurement reliability and reproducibility. Replicate phenotypes from an individual can be averaged, when appropriate. Multiple measurement modalities, for instance, quantifying carotid atherosclerosis using both ultrasound and magnetic resonance imaging, may also bolster phenotype accuracy and precision. <sup>13</sup> Overall, deep phenotyping should include reliable, comprehensive, and high-resolution assessment of the known components of the phenotype of interest.

**TABLE 2.** Phenomic Analysis Techniques

Techniques	<b>Example Methods</b>	Output
Physician assessment	Integrated clinical assessment from physician experience	Diagnosis
Heat maps	1- and 2-color maps of discrete and continuous data	Visual representation of multidimensional data
Clustering	Various clustering algorithms	Dendrogram, sorted heat maps
Text mining	Database and electronic medical record mining	Extracting concepts and relationships from text
Pathway analysis	Gene ontology, network analysis	Potential gene-subphenotype associations, gene-phenotype networks

## PHENOMIC ANALYSIS

Development of deep phenotypes creates difficult-to-interpret multidimensional data. Luckily, visualization and analysis techniques already exist for drawing conclusions from multidimensional data (Table 2). Remarkably, Sneath<sup>14</sup> described a computational method to catalog and score similarities between bacterial species to create a taxonomic classification more than 50 years ago. Eisen et al.<sup>15</sup> described a now widely used method, using heat maps and clustering to identify groups of coexpressing genes from highly multidimensional microarray expression data. These techniques can easily be adapted for the visual representation and analysis of human deep phenotypes.

The construction of a phenotype heat map consists of placing either the major phenotype classes or individual subjects in rows, and the quantitative or qualitative subphenotypes in columns of a grid. In a 1-color heat map, black and white boxes indicate the presence and absence, respectively, of an unambiguous discrete or qualitative phenotype or trait. For quantitative traits, grayscale can be used to indicate degree of affection. In a 2-color heat map, one color can be used to indicate a positive fold change from a normative value, whereas the other color indicates a negative fold change from the normative value; greater intensities indicate larger fold changes.

Once heat maps of quantitative or qualitative phenotype data are generated, cluster analysis can help group similar observations into subgroups for identifying potential relationships between data subsets. Software can assist in constructing and analyzing heat maps, such as Hierarchical Cluster Explorer (www.cs.umd.edu/hcil/hce/). 16 There are many types of cluster analysis, although biologists are likely most familiar with hierarchical cluster analysis from sequence and phylogenetic evaluations. 15 The output of hierarchical clustering is a dendrogram, or relationship tree, in which all observations are leaves and more closely related observations emanate from more proximal branch points. Hierarchical clustering is a sequential procedure that can be either agglomerative, which begins with the 2 closest leaves and then adds the next closest leaf, and so on, or divisive, which involves the iterative removal of the most distant leaf. 17 The major limitation of cluster analysis is its dependence on the similarity metric—a measure of correlationthat is used to calculate "closeness" between any 2 observations. For example, the similarity metric could include a weighting factor because 1 subphenotype measure may contribute more information toward the "closeness" of 2 major phenotypes. Hence, different weighting strategies could lead to different conclusions. Nonetheless, cluster analysis is a tool to extract solutions from complex multidimensional data, including phenomic data.

Text mining and pathway analysis strategies have also been proposed for further refining deep phenotypes and uncovering new gene-phenotype associations. <sup>12</sup> In addition, text mining techniques could be used for the development of phenotypes from electronic medical records. Many well-known databases, such as Online Mendelian Inheritance in Man (OMIM) and PhenoGO, already house phenotype- and gene-disease associations. <sup>12</sup> Gene otology (GO) and pathway analysis techniques, also borrowed from gene expression studies, could provide new insight into GWASs for hypothesis generation. <sup>18</sup>

## ADVANCING GENOMIC STUDIES THROUGH PHENOMICS

#### Implicating a Novel Disease Mechanism

We described Old Order Amish patients with a neonatal syndrome of endocrine gland hypoplasia, cerebral anomalies,

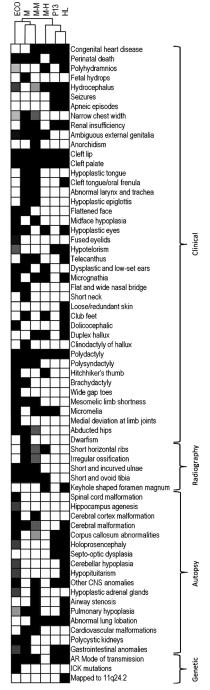


FIGURE 1. Phenomic analysis to identify a syndrome as novel. Phenomic analysis of 6 related congenital syndromes suggests that endocrine–cerebro-osteodysplasia (ECO) is a unique and distinct new syndrome. The heat map depicts clinical, radiographic, and autopsy phenotype information, as well as the results of genetic investigations. Each square is shaded from white (absent) to black (present), with intermediate gray shading indicating inconsistent presence. Hierarchical cluster analysis produced a dendrogram, the tree at the top of the figure, which illustrates the similarities between the syndromes. The additional syndromes studied include Majewski (M), Mohr-Majewski (M-M), Majewski-hydrolethalus (M-H), pseudotrisomy 13 (P13), and hydrolethalus (HL). AR indicates autosomal recessive; CNS, central nervous system; ICK, intestinal cell kinase gene.

and severe osteodysplasia. 19 Informed consent for study was provided by the parents of affected children, and the study was approved by the Office of Research Ethics at the University of Western Ontario. Initial examination of the affected infants suggested similarities to Majewski syndrome (OMIM: 263520) and hydrolethalus syndrome (OMIM: 236680). Collection of approximately 70 phenotype observations per patient and hierarchical cluster analysis indicated that the infants were affected with a novel disorder, subsequently named endocrine-cerebroosteodysplasia (Fig. 1), and further suggested that a novel molecular mechanism could be responsible. 19 Autozygosity mapping and targeted sequencing identified a rare mutation, proven through biochemical studies to be disease causing, in the gene encoding intestinal cell kinase. Thus, phenomic analysis to demonstrate both phenotype homogeneity among affected children and the presence of a constellation of phenotypes in a new syndrome of unknown etiology were important initial steps in this study.

In biomedicine, there are numerous examples of insight into normal human physiology gained from studying rare conditions. Similarly, studies of induced mutant mice have repeatedly shown the benefits of focusing on the phenotypic differences resulting from altered function or expression of a single gene. In humans, studies of naturally occurring rare diseases using linkage and autozygosity mapping strategies have become less common because attention has shifted toward common complex conditions. However, studies of well-defined rare syndromes still have their place, and with the reduced cost of high-density genotyping arrays, mapping studies are feasible in a laboratory with modest resources for even the rarest orphan disease. Furthermore, with the proliferation of next-generation

genomic sequencing efforts, phenomic analysis of mutation carriers will be essential to guide evaluation of the underlying pathophysiology resulting from a novel mutation, following the "reverse genetics" paradigm.

#### **Increasing Sample Homogeneity**

We have quantified adipose tissue depots in patients with lipodystrophy using magnetic resonance imaging to develop highly resolved phenotypes. <sup>21–23</sup> Using phenomics, such quantitative data have made clear the phenotypic distinctions between individuals with different genetic forms of familial partial lipodystrophy (FPLD), once considered to be a single entity (Fig. 2). <sup>24,25</sup> Familial partial lipodystrophy type 3 (FPLD3) has milder adipose tissue atrophy but more severe metabolic complications including insulin resistance, hypertension, dyslipidemia, and earlier-onset type 2 diabetes when compared with FPLD type 2 (FPLD2). <sup>25</sup> In addition, FPLD3 patients experience blunted response to thiazolidinediones compared with FPLD2 patients. <sup>26</sup>

Finding analogous phenotypic distinctions in common complex diseases could improve the signal-to-noise ratio for genotype-phenotype association analysis. Again, using cardio-vascular disease as an example, it has been suggested that the "distance" between underlying atherogenic mechanisms and disease end points may contribute to study heterogeneity. Through phenomics, the identification of patient subgroups with a specific mechanism leading to cardiovascular disease could improve the discriminatory power for genetic association studies, although making these distinctions and creating patient subgroups simultaneously reduces sample size.

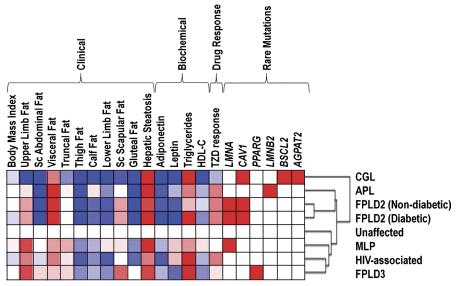


FIGURE 2. Phenomic analysis to identify subphenotypes. A 2-color heat map of human lipodystrophy syndromes demonstrates that, although clinically similar, careful phenomic analysis can clearly show phenotypic differences that may be the result of different molecular etiologies. The included syndromes are congenital generalized lipodystrophy (CGL), acquired partial lipodystrophy (APL), metabolic laminopathy (MLP), human immunodeficiency virus—associated lipodystrophy (HIV-associated), and FPLD2 and FPLD3. The fold change of severity or frequency was compared with normal reference range, with darker shades of red indicating increased severity or prevalence of the feature and darker shades of blue indicating decreased severity or prevalence of the feature. Hierarchical cluster analysis produced a dendrogram, which depicts the relationship between syndromes and indicates that FPLD2 and FPLD3, caused by mutations within the nuclear lamin A/C gene (LMNA) and peroxisome proliferator—activated receptor-gamma gene (PPARG), respectively, are clearly different by phenomic analysis. AGPAT2 indicates 1-acylglycerol-3-phosphate acetyltransferase 2 gene; BSCL2, Berardinelli-Seip congenital lipodystrophy type 2 gene; CAV1, caveolin-1 gene; HDL-C, high-density lipoprotein cholesterol; LMNB2, nuclear lamin B2 gene; TZD, thiazolidinedione.

# **Generating Molecular Hypotheses**

Pleiotropy, in which multiple disease phenotypes are caused by mutations in the same gene, is illustrated by the family of disorders called "laminopathies," which are due to a range of mutations within LMNA, which encodes nuclear lamin A/C.<sup>8</sup> Laminopathies include a diverse range of phenotypes including partial lipodystrophy, dilated cardiomyopathy, muscular dystrophy, and premature aging syndromes.8 Careful analysis of the phenotypes in carriers of LMNA mutations indicates some commonalities across this family of widely disparate disorders.<sup>28</sup> Application of phenomic heat mapping and hierarchical clustering analysis identified 2 main classes of laminopathies based on organ system involvement. The distribution of mutations across the lamin A/C domains was nonrandom across the 2 laminopathy classes, suggesting that mutation position relative to the nuclear localization signal domain may be an important determinant of the resultant complex phenotype.2

# Merging of Clinical, Biochemical, and Genomic Information for Diagnosis Refinement

Incorporating genetic, biochemical, and phenotypic information into a single analysis can further refine classic clinical phenotypes. For example, the Fredrickson hyperlipoproteinemia

(HLP) phenotypes are defined by the quality and quantity of plasma lipid subfractions after ultracentrifugation and have been used clinically for decades.<sup>29</sup> However, molecular genetics research has uncovered the molecular pathways underlying many of the HLP phenoypes.<sup>30</sup> For example, mutations in the LDL receptor (LDLR), proprotein convertase subtilisin/kexin-type 9 (PCSK9), and apolipoprotein B (APOB) can all produce HLP type 2A, each by impairing different steps in the LDL cholesterol metabolism pathway.<sup>30</sup> Both rare mutations, such as those in genes encoding lipoprotein lipase (LPL) and apolipoprotein C2 (APOC2), and common single-nucleotide polymorphisms, such as those in glucokinase regulatory protein (GCKR) and tribbles homolog 1 (TRIB1), contribute to overall HLP susceptibility. <sup>31,32</sup> Genomic resequencing and association studies show that severe hypertriglyceridemia (HLP type 5) is a mosaic of both common and rare genetic variants with a wide range of effect sizes (Fig. 3). 30-33 Perhaps the traditional classification of lipid phenotypes will need to be readdressed in light of discoveries into the disease-causing mechanisms gleaned from genetic investigations. Knowledge of the exact diseasecausing mechanism in a specific patient from clinical, biochemical, and genomic investigations may lead to a refinement of diagnosis and identify patients who are most likely to benefit from a specific pharmacological intervention.

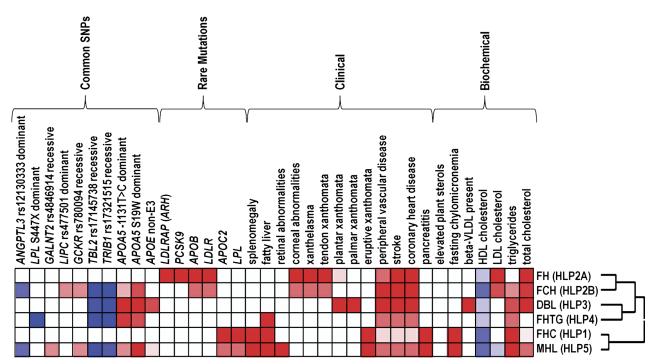


FIGURE 3. Phenomic analysis to refine diagnosis. A 2-color heat map contrasts the similarities and differences between the Fredrickson HLP phenotypes (HLP1-HLP5) and visually represents the contribution of both common single-nucleotide polymorphisms and rare mutations. Defining features are listed in columns and HLP phenotypes are in rows. The fold change of severity or presence was compared with normal reference range, with darker shades of red indicating increased severity or prevalence of the trait and darker shades of blue indicating decreased severity or prevalence of the trait. Effect size of common variants calculated from genotypes of 386 HLP patients collected at a tertiary referral lipid clinic.<sup>31</sup> All patients provided informed consent for research participation and genomic analysis, and the study was approved by the Office of Research Ethics at the University of Western Ontario. ANGPTL3 indicates angiopoietin-like 3; APOA5, apolipoprotein A5; APOB, apolipoprotein B; APOC2, apolipoprotein C-II; APOE, apolipoprotein E; ARH, autosomal recessive hypercholesterolemia; DBL, dysbetalipoproteinemia; FCH, familial combined hyperlipidemia; FHC, familial hyperchylomicronemia; FH, familial hypercholesterolemia; FHTG, familial hypertriglyceridemia; GALNT2, N-acetylgalactosaminyl transferase 2; GCKR, glucokinase regulatory protein; HDL, high-density lipoprotein; LDLR, low-density lipoprotein receptor; LDLRAP, LDL receptor adaptor protein; LIPC, hepatic lipase; LPL, lipoprotein lipase; MHL, mixed hyperlipidemia; TBL2, transducin β-like 2; TRIB1, tribbles homolog 1.

# Phenomics and Mendelian Randomization

Mendelian randomization (MR) uses the random assortment of alleles at meiosis to evaluate a causal relationship between an intermediate biomarker and a disease end point. 34,35 If a genetic variant is associated with the circulating concentration of a biomarker, and the biomarker is causal for the disease end point, the genetic variant should also be associated with the disease end point. There are several caveats to the interpretation of MR studies, but in theory, MR can provide support for causal relationships.<sup>36</sup> Efforts to curate databases of genotype-phenotype associations are already underway for identifying networks of interrelated findings. 12 Even if a genetic variant-phenotype association does not surpass the stringent significance threshold required in GWASs owing to multiple testing, confidence would be increased if the genetic variant is consistently associated with multiple intermediate steps in a particular pathogenic pathway. 18 A network of intermediate steps could be characterized in the living study participant through deep phenotyping and phenomic analysis.

#### **SUMMARY**

Owing to recent technological advances, the bar has been raised for data quality and quantity in genetic association studies. To tease out the small genetic effects operating in complex diseases, clinicians will be required to collect precise deep phenotypes to reduce measurement error, reduce study heterogeneity, and further refine study populations. Through analysis of individuals with rare mutations identified through next-generation sequencing, phenomics will play an important role in hypothesis generation. Phenomics will create new diagnostic criteria through the integration of clinical, diagnostic, and genetic information. Finally, confidence in the biological meaning of genetic associations will be improved by assessing the genetic variants effect on multiple components of a complex pathway.

# **ACKNOWLEDGMENTS**

The authors thank Dr. Tisha Joy for providing data included in Figure 2. R.A.H. is a Career Investigator of the Heart and Stroke Foundation of Ontario, holds the Edith Schulich Vinet Canada Research Chair (Tier I) in Human Genetics, the Martha G. Blackburn Chair in Cardiovascular Research, and the Jacob J. Wolfe Distinguished Medical Research Chair at the University of Western Ontario.

#### **REFERENCES**

- Willer CJ, Speliotes EK, Loos RJ, et al. Six new loci associated with body mass index highlight a neuronal influence on body weight regulation. Nat Genet. 2009;41:25–34.
- 2. Freimer N, Sabatti C. The human phenome project. *Nat Genet*. 2003;34:15–21.
- Schork NJ. Genetics of complex disease: approaches, problems, and solutions. Am J Respir Crit Care Med. 1997;156:S103–S109.
- 4. Scriver CR. After the genome—the phenome? *J Inherit Metab Dis*. 2004;27:305–317.
- Lander ES, Schork NJ. Genetic dissection of complex traits. Science. 1994;265:2037–2048.
- Wong MY, Day NE, Luan JA, et al. The detection of gene-environment interaction for continuous traits: should we deal with measurement error by bigger studies or better measurement? *Int J Epidemiol*. 2003;32:51–57.

- Teo YY, Inouye M, Small KS, et al. A genotype calling algorithm for the Illumina BeadArray platform. *Bioinformatics*. 2007;23: 2741–2746.
- 8. Hegele RA, Oshima J. Phenomics and lamins: from disease to therapy. *Exp Cell Res.* 2007;313:2134–2143.
- 9. Burton PR, Hansell AL, Fortier I, et al. Size matters: just how big is BIG?: quantifying realistic sample size requirements for human genome epidemiology. *Int J Epidemiol*. 2009;38:263–273.
- Plomin R, Haworth CM, Davis OS. Common disorders are quantitative traits. *Nat Rev Genet*. 2009;10:872–878.
- Tracy RP. 'Deep phenotyping': characterizing populations in the era of genomics and systems biology. Curr Opin Lipidol. 2008;19: 151–157.
- Lussier YA, Liu Y. Computational approaches to phenotyping: high-throughput phenomics. *Proc Am Thorac Soc.* 2007;4: 18–25
- Spence JD, Hegele RA. Noninvasive phenotypes of atherosclerosis: similar windows but different views. Stroke. 2004;35:649–653.
- Sneath PH. The application of computers to taxonomy. J Gen Microbiol. 1957;17:201–226.
- Eisen MB, Spellman PT, Brown PO, et al. Cluster analysis and display of genome-wide expression patterns. *Proc Natl Acad Sci* USA. 1998;95:14863–14868.
- Seo J, Shneiderman B. Knowledge discovery in high-dimensional data: case studies and a user survey for the rank-by-feature framework. *IEEE Trans Vis Comput Graph*. 2006;12:311–322.
- Sherlock G. Analysis of large-scale gene expression data. Curr Opin Immunol. 2000;12:201–205.
- Torkamani A, Schork NJ. Pathway and network analysis with high-density allelic association data. *Methods Mol Biol*. 2009;563: 289–301.
- Lahiry P, Wang J, Robinson JF, et al. A multiplex human syndrome implicates a key role for intestinal cell kinase in development of central nervous, skeletal, and endocrine systems. *Am J Hum Genet*. 2009;84:134–147.
- Peltonen L, Uusitalo A. Rare disease genes—lessons and challenges. Genome Res. 1997;7:765–767.
- Al-Attar SA, Pollex RL, Robinson JF, et al. Semi-automated segmentation and quantification of adipose tissue in calf and thigh by MRI: a preliminary study in patients with monogenic metabolic syndrome. *BMC Med Imaging*. 2006;6:11.
- Al-Attar SA, Pollex RL, Robinson JF, et al. Quantitative and qualitative differences in subcutaneous adipose tissue stores across lipodystrophy types shown by magnetic resonance imaging. BMC Med Imaging. 2007;7:3.
- Hegele RA, Joy TR, Al-Attar SA, et al. Thematic review series: adipocyte biology. Lipodystrophies: windows on adipose biology and metabolism. *J Lipid Res.* 2007;48:1433–1444.
- Hegele RA, Pollex RL. Genetic and physiological insights into the metabolic syndrome. Am J Physiol Regul Integr Comp Physiol. 2005;289:R663–R669.
- Joy T, Hegele RA. Genetics of metabolic syndrome: is there a role for phenomics? Curr Atheroscler Rep. 2008;10:201–208.
- Hegele RA. Monogenic forms of insulin resistance: apertures that expose the common metabolic syndrome. *Trends Endocrinol Metab*. 2003;14:371–377.
- Manolio TA, Boerwinkle E, O'Donnell CJ, et al. Genetics of ultrasonographic carotid atherosclerosis. *Arterioscler Thromb Vasc Biol.* 2004;24:1567–1577.
- 28. Hegele R. *LMNA* mutation position predicts organ system involvement in laminopathies. *Clin Genet.* 2005;68:31–34.

- Fredrickson DS, Lees RS. A system for phenotyping hyperlipoproteinemia. *Circulation*. 1965;31:321–327.
- Hegele RA. Plasma lipoproteins: genetic influences and clinical implications. Nat Rev Genet. 2009;10:109–121.
- Hegele RA, Ban MR, Hsueh N, et al. A polygenic basis for four classical Fredrickson hyperlipoproteinemia phenotypes that are characterized by hypertriglyceridemia. *Hum Mol Genet*. 2009;18:4189–4194.
- Wang J, Ban MR, Kennedy BA, et al. APOA5 genetic variants are markers for classic hyperlipoproteinemia phenotypes and hypertriglyceridemia. Nat Clin Pract Cardiovasc Med. 2008;5: 730–737.
- Wang J, Cao H, Ban MR, et al. Resequencing genomic DNA of patients with severe hypertriglyceridemia (MIM 144650). Arterioscler Thromb Vasc Biol. 2007;27:2450–2455.
- 34. Davey Smith G, Ebrahim S. 'Mendelian randomization': can genetic epidemiology contribute to understanding environmental determinants of disease? *Int J Epidemiol*. 2003;32:1–22.
- Thanassoulis G, O'Donnell CJ. Mendelian randomization: nature's randomized trial in the post-genome era. *JAMA*. 2009;301: 2386–2388.
- Sheehan NA, Didelez V, Burton PR, et al. Mendelian randomisation and causal inference in observational epidemiology. *PLoS Med*. 2008;5:e177.