The Measurement of 3-Epimer 25-Hydroxyvitamin D by Mass Spectrometry in Clinical Specimens Detects Inconsequential Levels in Adult Subjects

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Background: Vitamin D is derived from dietary sources or from the action of ultraviolet light on 7-dehydrocholesterol and undergoes a number of enzymatic modifications that lead to the synthesis of active vitamin D metabolites or metabolites with reduced biological activity. Among these, epimerization at the 3-hydroxyl group leads to the synthesis of 3-epimer 25-hydroxyvitamin D (3EVD). Described first in biological system experiments using in vitro incubation of vitamin D in cell culture, this molecule has been reported as having distinct activities when compared with 25-hydroxy vitamin D (25OHVD). Measurements of vitamin D have been conducted using a variety of methodologies and have led to conflicting assessments of the quantities of 3EVD₃ that are measured.

Method: The present article describes the development and use of a simple liquid chromatography–tandem mass spectrometry method validated by the Clinical Laboratory Improvement Amendments to quantitate 3EVD₃ in 3528 subjects, including 309 children (162 are <2 years) and 232 pregnant women.

Results: Our findings demonstrate that, although 3EVD_3 constitutes a significant proportion of measureable 25OHVD_3 in subjects younger than 1 year, 3EVD_3 levels are negligible in most subjects older than 1 year.

Conclusions: It is important to choose the correct 25OHVD assay dependent on the age of the patient. Patients younger than 1 year should be run on a liquid chromatography-tandem mass spectrometry assay proven to not have potential contributions from any 3EVD present in the sample.

Key Words: 25-hydroxy vitamin D, mass spectrometry, 3-epimer, pediatric, quantitative assay, adult, CLIA-validated assay

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Vitamin D has long been recognized to play a critical role in the development and maintenance of bone mineral density. Originally identified in the context of its role in the prevention and treatment of rickets, it has been recognized more recently that subtler forms of vitamin D deficiency are prevalent and represent a major factor in the development of osteopenia and osteoporosis in the adult population.^{1–3} Furthermore, a number of studies have implicated vitamin D deficiency as

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contributing to the pathogenesis or progression of cardiovascular disease, cancer, multiple sclerosis, and abnormalities of the immune system.^{4–9}

Vitamin D is derived from dietary sources (vitamin D_2 or D_3) or from the action of ultraviolet light on 7-dehydrocholesterol in the skin to form cholecalciferol (vitamin D_3). After transport to the liver, vitamin D_3 is hydroxylated at the 25 position by CYP2R1 to form 25-hydroxyvitamin D_3 (25OHVD₃).¹⁰⁻¹³ Hydroxylation at the 1 position of 25OHVD occurs predominantly in the kidney and is responsible for the synthesis of the most active form of vitamin D, 1,25-dihydroxyvitamin D,¹⁴ which exerts its effects by binding to the vitamin D receptor, a member of the nuclear receptor family.¹⁵

In addition to processes leading to the synthesis of active forms of vitamin D, additional enzymatic steps have been identified that further modify vitamin D. Hydroxylation at the 24 position leads to the synthesis of 24,25-dihydroxyvitamin D. The synthesis of 24,25-dihydroxyvitamin D is stimulated by increased levels of 1,25-dihydroxyvitamin D, and this enzymatic conversion is felt to serve as a mode of regulating circulating levels of 1,25-dihydroxyvitamin D. The 24,25-dihydroxyvitamin D lacks prototypical activity as vitamin D in many assays but may serve alternate roles in vitamin D signaling.^{16,17}

In like fashion, the hydroxyl group at the C-3 position of vitamin D may be configured either in an R or S configuration. The formation of 3-epimer (3-epi) 25-hydroxyvitamin D (3EVD), which has the hydroxyl group at the 3 position in the S configuration, was originally identified in incubation studies using cultured neonatal keratinocytes but has subsequently been detected in a number of tissues and in clinical samples.^{18–25} Al-though 3EVD can be converted to 3-epi 1,25-dihydroxyvitamin D, 3-epi 1,25-dihydroxyvitamin D lacks activity in some biological assays of vitamin D action.^{26–28} As such, most recent attention has been focused on 3EVD as it pertains to its role as a potential confounder to the accurate determination of 25OHVD.

The recognition of the prevalence of vitamin D deficiency in the general population has resulted in a dramatic rise in the number of tests to measure 25OHVD that are performed annually in the United States and worldwide.²⁹ In concert with the rise in the numbers of tests performed is the recognition that tests to measure 25OHVD remain poorly standardized and that carefully characterized reference standards are essential.^{30–32}

The recent application of liquid chromatography and tandem mass spectrometry (LC-MS/MS) has permitted the increasingly precise measurement of 25OHVD levels. Along with this precision, however, is the recognition that specific metabolites of vitamin D may not be measured owing to their distinct chemical properties. Finally, the application of other methods to the measurement of 25OHVD levels has led to some uncertainty as to whether metabolites, such as 3EVD₃, constitute an important potential confounder to the measurement of vitamin D levels in distinct patient groups.

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 $3EVD_2$ and $3EVD_3$ epimers. The assay was shown to be rapid and accurate, with a lower limit of quantitation (LOQ) of 3 ng/mL

for 25OHVD₂, 25OHVD₃, 3EVD₂, and 3EVD₃. The data ac-

quired also demonstrated the robustness of the assay and its

reproducibility with coefficients of variation of 8% to 13% at

the 3 concentration levels (15, 40, and 80 ng/mL) analyzed for

3EVD was not separated) and the new methodology reported

in the present article (which permits the chromatographic sep-

aration and measurement of 3EVD levels). As depicted in

Figure 1, these measurements showed excellent agreement. Supplemental Figures 1 and 2 (Supplemental Digital Content 1,

http://links.lww.com/JIM/A20) show a bias plot and histogram,

respectively, of the same analysis, providing further proof of the

Levels of 3EVD₃ Are Infrequently Measureable in

3EVD₃ revealed levels of 25OHVD₂ greater than the limit

of detection (1 ng/mL) in only a small number of individuals

(n = 2), with no samples containing levels of $3EVD_2$ greater

sureable 25OHVD₃ were highly variable, ranging from 4 to

126 ng/mL. In the 2987 specimens analyzed from subjects

18 years or older, only 65 (2.2%) had measureable levels of

3EVD₃ (>3 ng/mL), with only 15 (0.47%) having values of ng/mL

The measurement of 25OHVD₂, 25OHVD₃, 3EVD₂, and

The levels of 25OHVD₃ in samples containing mea-

Specimens From Subjects 18 Years or Older

than the lower LOQ (3 ng/mL) in this age group.

excellent agreement between the 2 protocols.

A direct comparison of the values of 25OHVD was conducted in an adult subset constituting 2987 patients older than 18 years using both the existing assay for 25OHVD (in which

all analytes.

or greater (Fig. 2).

MATERIALS AND METHODS

Standards/Calibrators, Internal Standard, and Controls

Standards for $3EVD_3$, $3EVD_2$, $25OHVD_3$, and $25OHVD_2$ were purchased from Isosciences (King of Prussia, PA). The calibrators and controls were spiked into vitamin D metabolite depleted diluent from SeraCare Life Sciences (Milford, MA).

Study Samples

Three different populations of samples were analyzed. Discarded samples submitted for the clinical analysis of 25OHVD were subjected to the measurement of $3EVD_2$ and $3EVD_3$, 25OHVD₂, and 25OHVD₃ using the assay described in this article. The analysis of discarded specimens was judged exempt by the Western Institutional Review Board. All pediatric samples and third trimester of pregnancy samples were obtained from patient discards 3 weeks after results had been provided to the ordering physician. Discarded samples from women in their first and second trimesters of pregnancy were purchased from Biotheme Research Solutions.

Sample Preparation

In both of the LC-MS/MS assays used in this study, patient samples were extracted using a direct-in-plate protein precipitation and filtration. This step was automated on a Perkin-Elmer Janus robot by adding 100 μ L of patient serum to each well of a 96-well protein precipitation plate with a regular deep-well collection plate underneath it. The Impact protein precipitation plate from Phenomenex (Torrance, CA), together with the collection plate, was transferred to the Agilent Bravo System to dispense 400 μ L of methanol containing the internal heavy isotope-labeled standard (D₃-250HVD₃) to each well. It was then vortexed and spun down in a centrifuge until all supernatant settled in the collection plate and was then injected onto the LC-MS/MS system.

Liquid Chromatography-MS/MS

A Cohesive Technologies Aria TLX-4 System (ThermoFisher, San Jose, CA) was used for LC with separation accomplished using a 100 \times 2.1-mm, 2.6-µm pentafluorophenyl solid-core high-performance LC column (Accucore from ThermoFisher Scientific, San Jose, CA). The mobile phase solvents were 0.1% aqueous formic acid and methanol, which were delivered in a gradient from 70% to 90% methanol during a period of 6 minutes 15 seconds (including column equilibration time).

Thermo Scientific TSQ Quantum Ultra MS/MS System with atmospheric pressure chemical ionization source was used as the detector for both assays used in this study. Mass spectrometer data were obtained in positive ion mode. Peak area ratios of the analytes and internal standards were used for calculations of concentrations. The precursor ions and its major fragment ions (*m/z*) are as follows: $395.3 \rightarrow 179.1 + 209.1 + 251.1$ for $250HVD_2$ and $3EVD_2$ and $383.3 \rightarrow 159.1 + 211.1$ for $250HVD_3$ and $3EVD_3$. The same fragments were used to measure $250HVD_2$ and $250HVD_3$ in both assays used in this study.

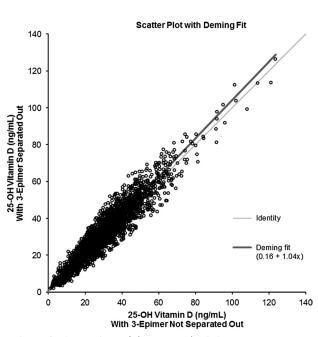
RESULTS

Assay Performance and Correlation With Existing Assay

The data collected during the Clinical Laboratory Improvement Amendments validation established a method for the determination of $250HVD_3$ and $250HVD_2$ as well as the

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FIGURE 1. Comparison of the new and existing assays to measure 25OHVD. Individual adult samples (n = 2987) were assayed to measure 25OHVD using either the existing LC/MS method (*x* axis) or the method described in the present article (*y* axis), in which 3EVD is chromatographically separated from 25OHVD, permitting a quantitation of 25OHVD, without any potential contribution of 3EVD.



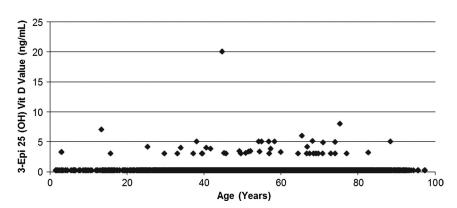


FIGURE 2. Absolute 3EVD values in subjects 2 to 97 years old (n = 3134).

Pediatric Samples Have an Increased Proportion of Samples With Measureable Levels of 3EVD₃, Which Declines With Age

In contrast to results obtained using samples obtained from older subjects, levels of 3EVD_3 were measureable at a much higher frequency in the 309 pediatric specimens tested (<18 years old). This statement relates not only to the absolute levels of 3EVD_3 detected (Fig. 3) but also to the frequency with which samples contained measureable 3EVD_3 (supplemental Figure 3, http://links.lww.com/JIM/A20). Comparison of Figure 3 and Supplemental Figure 3, http://links.lww.com/JIM/A20, demonstrates that the proportion of samples in which 3EVD_3 is detectable is less than 15% after age of 1 year (with that percentage dropping with increasing age) and that the concentrations measured in subjects older than 1 year are near the limits of detection (3 ng/mL; Table 1).

Pregnancy

The demonstration that young children have a higher incidence of measureable levels of 3EVD and that, when measureable, have values that are more frequently substantial prompted us to examine the measured levels of 3EVD in pregnant women at various stages in their pregnancy. Of the 232 samples assayed, 44 of 232 subjects (19%) had detectable 3EVD₃ concentrations (mean, 5.1 ng/mL; range, 3–8). Notably, although these levels are low and relatively infrequent, 24 of

these measurements were in individuals with low levels of measured $25OHVD_3$ (5 ng/mL of $3EVD_3$ with 35 ng/mL of $25OHVD_3$; 8 ng/mL of $3EVD_3$ with 35 ng/mL of $25OHVD_3$; 3 ng/mL of $3EVD_3$ with 20 ng/mL of $25OHVD_3$; 3 ng/mL of $3EVD_3$ with 25 ng/mL of $25OHVD_3$), in which the levels of $3EVD_3$ detected could cloud the interpretation of the measured $25OHVD_3$ concentrations.

Relationship Between Levels of 3EVD and Levels of 25OHVD

Although the bulk of our attention was drawn to the changes that are evident in the different populations that we examined (young pediatric subjects, older pediatric subjects, adults, pregnant women), we noted that the proportion of $3EVD_3$ detected exhibited a crude, inverse relationship to the measured levels of $25OHVD_3$. The largely unchanging levels of measured $3EVD_3$ would support that this relationship is a reflection of changes in the measured $25OHVD_3$. These relationships are depicted in Figures 4A and B.

DISCUSSION

These data establish a rapid method for the separation and measurement of $250HVD_3$ and $250HVD_2$ along with their epimers $3EVD_3$ and $3EVD_2$. This assay is rapid and accurate with an LOQ of 3 ng/mL. The data presented here demonstrate the robustness of the assay and its reproducibility with

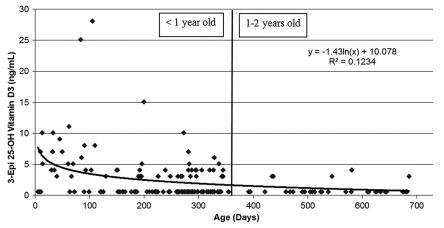


FIGURE 3. Absolute level of epimer D_3 in children younger than 2 years (n = 162). The absolute level of $3EVD_3$ measured (y axis) is plotted versus the age of the subject (in days) on the x axis. Although measurable at a low level in children between 1 and 2 years old, all measured levels of 3-epi were less than 4 ng/mL.

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	Measurable/ Analyzed, n (%)	Mean of Measureable Values, ng/mL	Range, ng/mL
<2 mo	10/13 (77)	6.6	3-10
2 and 3 mo	11/15 (73)	17.9	3-28
4 and 5 mo	3/12 (25)	3.6	3–4
6 and 7 mo	8/20 (40)	5	3-15
8 and 9 mo	13/39 (33)	4.3	3-10
10 and 11 mo	9/29 (31)	4	3–6
12–24 mo	6/34 (17)	3.1	3–4
24-36 mo	5/26 (19)	3	3–3
36–48 mo	2/19 (10)	3	3–3
48–60 mo	2/37 (5)	3.5	3–4
60–72 mo	1/21 (5)	3	3–3
72–84 mo	4/44 (9)	4	3–5

TABLE 1. Distribution of Measureable Values of $3EVD_3$ in Subjects Aged 0 to 6 Years

coefficients of variation of 8% to 13% for 25OHVD and 3EVD in the 3 concentration ranges tested.

Application of this assay demonstrates that 3EVD_3 and 3EVD_2 concentrations were measureable in only 65 of 3134 samples (2.2%) obtained from nonpregnant subjects 2 years and older (Fig. 2). This low frequency of measurable values is paralleled by the consistently lower levels detected in this group,

with only 18 samples (0.57%) having measureable values of

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5 ng/mL or greater. Further breakdown of this group to adults only (\geq 18 years old, n = 2987) showed that, although most values of 5 ng/mL or greater (n = 15) were in this age range, this did not affect the rate of positivity (0.47%) because the bulk of the patients fell in this age range.

The distribution of results from adult patients was in stark contrast to the measurements performed in specimens obtained from the younger subjects. The frequency of measureable levels of 3EVD was observed to be substantially higher in subjects younger than the 2 years (Fig. 3). In this age range, both the frequency of measureable values and the absolute levels of 3EVD measured increased significantly. These findings are broken down by chronological age of the pediatric patients in Table 1. Of note, although measurable at a low frequency in children older than 1 year, all measured levels of 3EVD were less than 5 ng/mL in subjects older than 1 year.

Finally, in pregnancy, it was noted that a significant portion of the samples tests (44/232, 19%) had detectable levels of 3EVD with the mean value being 5.1 ng/mL. However, 24 of those 44 samples (24/232, 10%) had values high enough that they could affect the clinical definition of the patient's total vitamin D status. Because of this, it may be advisable to use a test such as the one detailed in this report to measure vitamin D status in pregnant women to ensure accuracy.

The biological importance of 3EVD remains to be clarified. Although 3-epi 1,25-dihydroxyvitamin D has been shown to be capable of modulating some biological effects of vitamin D, it does not possess all attributes associated with 1,25-dihydroxyvitamin D. As a result, the focus of most

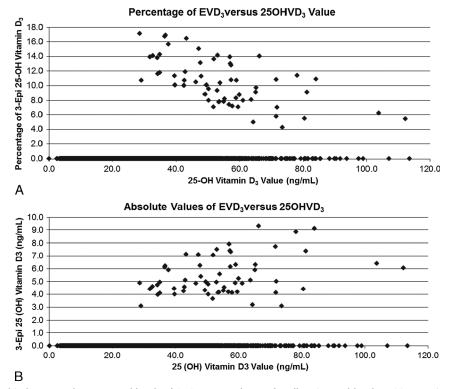


FIGURE 4. Relationship between the measured levels of $25OHVD_3$ and EVD_3 for all patients older than 18 years (n = 2987). Measured levels of $3EVD_3$ are displayed on the *y* axis as a percentage of the measured $25OHVD_3$ (shown on the *x* axis). In this representation, most individuals have levels of EVD_3 below the limits of assay detection. An inverse relationship is evident between the levels of $25OHVD_3$ and the percentage of EVD_3 in individuals in whom EVD_3 levels are measureable. Figure 4B shows the same representation, but as absolute values rather than percentages.

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investigations relating to 3EVD has centered on the recognition that this molecule could represent a significant confounder in the measurement of $250HVD_3$. This issue has been examined in a number of studies.

Singh et al.³³ used an LC/MS assay to examine levels of 3EVD in 439 individuals, including 183 individuals younger than 1 year and 47 individuals aged 1 to 18 years. This group demonstrated that subjects younger than 1 year often had measureable levels of 3EVD. In this series, 3EVD was not detected in patients older than 1 year (n = 256).

In a limited study, Stepman et al.³⁴ used an ultra performance liquid chromatography-MS/MS assay to examine specimens from 6 infants aged between 1 and 4 months and 32 samples from adults. This analysis detected 3EVD in all 6 infants, comprising 15% to 44% of the 25OHVD measured. The 3EVD₃ was detected in all 32 adult specimens assayed, but at lower levels compared with the infant specimens (between 2.5% and 17% of the 25OHVD₃ measured).

In a letter to the editor, van den Ouwelanc and colleagues³⁵ used an LC-MS/MS method to separate and quantitate the levels of $3EVD_3$ in 51 subjects younger than 1 year, 74 subjects aged 1 to 10 years, and 104 individuals 18 years or older. These investigators were able to detect $3EVD_3$ in all sera from infants and in three fourths of adult samples. In this assay, the proportion of the total $25OHVD_3$ contributed by $3EVD_3$ ranged from a median of 11.1% in the youngest group, to 6.2% in the children, and to 3.5% in the adults.

Schleicher et al.³⁶ described an ultra performance liquid chromatography-MS/MS method for simultaneous measurement of 25OHVD₃, 25OHVD₂, and 3EVD₃ in human serum. These investigators applied this method to 2 convenience samples composed of 98 blood donors and 35 pregnant women. In these samples, 3EVD₃ was detectable in 43% of random blood donors and 80% of pregnant women. Of note, these investigators suggested that the concentrations of 3EVD₃ were correlated with the concentrations of 25OHVD₃.

Strathmann et al.³⁷ measured concentrations of 25OHVD₃ and 3EVD₃ in 125 subjects (aged 3 months to 1 year) and 626 patients aged 1 to 94 years undergoing routine 25OHVD testing. These investigators found higher proportion of 3EVD₃ in the youngest age groups with a trend to decreasing proportion of 3EVD₃ compared with the total with increasing age, which they attributed to constant levels of 3EVD₃ in the face of gradually increasing 25OHVD₃. Of note, their analysis suggests that ~24% of children and 3% of adults would be misclassified on the basis of the bias introduced by the measured levels of 3EVD₃.

Lensmeyer et al.³⁸ reported the application of an LC-MS/MS to measure the levels of $25OHVD_3$ and $3EVD_3$ in 214 clinical specimens from neonates to subjects 80 years and older. The $3EVD_3$ epimer was detected in 212 of 214 (99%) of samples. Concentrations ranged from 0.1 to 23.7 ng/mL for $3EVD_3$. Of note, in this assay, a clear relationship of $3EVD_3$ to $25OHVD_3$ was noted, with higher amounts of $3EVD_3$ measured in samples with higher $25OHVD_3$ concentrations.

Baecher et al.³⁹ reported the application of a highperformance LC-MS/MS method to measure $25OHVD_3$, $25OHVD_2$, $3EVD_3$, and $24R_25$ -dihydroxyvitamin D₃ in human serum. In these specimens (no clinical data available), the concentration of $3EVD_3$ was, on the average, approximately 7% (ranging from 4% to 20%) of the concentration of $25OHVD_3$.

Bailey et al.⁴⁰ used an LC-MS/MS method to measure $250HVD_3$ and $3EVD_3$. In all subjects, $3EVD_3$ was measured, with higher concentrations observed in infants. Within the first year of life, $250HVD_3$ concentrations increased linearly, whereas

 $3EVD_3$ concentrations remained constant. At 12 months old, $3EVD_3$ concentration dropped by almost 50%.

The results of the current study support and extend various items identified in prior reports.

First, our findings indicate that the proportion of subjects with measureable levels of 3EVD_3 is greatest at ages younger than 1 year.

Second, this larger sample set permits a clearer definition of the changes that occur at older ages: the proportion of subjects with measurable levels decreasing progressively and the absolute levels progressively declining.

Third, the high concordance seen between the currently offered 25OHVD LC-MS/MS assay and the method detailed in this article, coupled with the low prevalence of detectable 3EVD concentrations in the general population studied, gives further evidence of the negligible contribution of either 3EVD to the currently reported 25OHVD LC-MS/MS assays.

Finally, in contrast to the report of Lensmeyer, a crude inverse relationship between the levels of $250HVD_3$ and the levels of $3EVD_3$ was apparent in the patient population analyzed.³⁸

Overall, the results of this study reinforce the notion that, in very young pediatric patients (<1 year old), it is important to account for the potentially high level of 3EVD₃ that may be present in this specific age group. To that end, we have developed a highly selective and specific mass spectrometrybased assay that does not experience interference posed by this vitamin D metabolite. Furthermore, the highly powered analysis of the age range of 2 to 97 years shows negligible contributions to the overall 25OHVD measurement, whether the 3-epi is separated chromatographically by the LC-MS/MS assay. From these data, it would therefore seem advisable for ordering physicians to consider the use of an LC-MS/MS method capable of separating the 3-epi metabolite away from the 25OHVD analytes in pediatric patients younger than 1 year. On the basis of our study, the same concern does not seem warranted for patients older than 1 year.

REFERENCES

- 1. Holick MF. Vitamin D deficiency. N Engl J Med. 2007;357:266-281.
- Holick MF. The vitamin D deficiency pandemic and consequences for nonskeletal health: mechanisms of action. *Mol Aspects Med.* 2008;29:361–368.
- Holick MF. Vitamin D: a d-lightful solution for health. J Investig Med. 2011;59:872–880.
- Adorini L, Penna G. Control of autoimmune diseases by the vitamin D endocrine system. Nat Clin Pract Rheumatol. 2008;4:404–412.
- Bouillon R, Eelen G, Verlinden L, et al. Vitamin D and cancer. J Steroid Biochem Mol Biol. 2006;102:156–162.
- Plum LA, DeLuca HF. Vitamin D, disease and therapeutic opportunities. Nat Rev Drug Discov. 2010;9:941–955.
- Battault S, Whiting SJ, Peltier SL, et al. Vitamin D metabolism, functions and needs: from science to health claims. *Eur J Nutr.* 2013;52:429–441.
- Aranow C. Vitamin D and the immune system. J Investig Med. 2011;59:881–886.
- Li YC. Molecular mechanism of vitamin D in the cardiovascular system. J Investig Med. 2011;59:868–871.
- Cheng JB, Levine MA, Bell NH, et al. Genetic evidence that the human CYP2R1 enzyme is a key vitamin D 25-hydroxylase. *Proc Nat Acad Sci U S A*. 2004;101:7711–7715.

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- utation 25_hydrovuuitamin D. is matabalized to 2 and 25 bus
- Levine MA, Dang A, Ding C, et al. Tropical rickets in Nigeria: mutation of the CYP2R1 gene encoding vitamin D 25-hydroxylase as a cause of vitamin D dependent rickets. *Bone*. 2007;40:S60–S61.
- Dong Q, Miller WL. Vitamin D 25-hydroxylase deficiency. Mol Genet Metab. 2004;83:197–198.
- Al Mutair A, Nasrat GH, Russell DW. Mutation of the CYP2R1 vitamin D 25-hydroxylase in a Saudi Arabian family with severe vitamin D deficiency. J Clin Endocrin Metab. 2012;97:E2022–E2025.
- Kato S, Yanagisawa J, Murayama A. The importance of 25-hydroxyvitamin D₃ 1α-hydroxylase gene in vitamin D-dependent rickets. *Curr Opin Nephrol Hypertens*. 1998;7:377–383.
- Carlberg C, Campbell MJ. Vitamin D receptor signaling mechanisms: integrated actions of a well-defined transcription factor. *Steroids*. 2013;78:127–136.
- Makin G, Lohnes D, Byford V, et al. Target cell metabolism of 1,25-dihydroxyvitamin D₃ to calcitroic acid: evidence for a pathway in kidney and bone involving 24-oxidation. *J Biochem*. 1989;262:173–180.
- Reddy GS, Tserng KY. Calcitroic acid, end product of renal metabolism of 1,25-dihydroxyvitamin D₃ through C-24 oxidation pathway. *Biochemistry*. 1989;28:1763–1769.
- Reddy GS, Muralidharan KR, Okamura WH, et al. Metabolism of 1a,25-dihydroxyvitamin D₃ and one of its A-ring diastereomer 1a,25-dihydroxy-3-epivitamin D₃ in neonatal human keratinocytes. In: Norman AW, Bouillon R, Thomasset M, eds. *Vitamin D A Pluripotent Steroid Hormone: Structural Studies, Molecular Endocrinology and Clinical Applications.* New York, NY: Walter de Gruyter; 1994:172–173.
- Reddy GS, Siu-Caldera M-L, Schuster I, et al. Target tissue specific metabolism of 1a,25-dihydroxyvitamin D₃ through A-ring modification. In: Norman AW, Bouillon R, Thomasset M, eds. *Vitamin D, Chemistry, Biology and Clinical Applications of the Steroid Hormone*. Riverside, CA: University of California; 1997:139–146.
- Reddy GS, Muralidharan KR, Okamura WH, et al. Metabolism of 1alpha,25-dihydroxyvitamin D(3) and its C-3 epimer 1alpha,25-dihydroxy-3-epi-vitamin D(3) in neonatal human keratinocytes. *Steroids*. 2001;66:441–450.
- Brown AJ, Ritter C, Slatopolsky E, et al. 1a,25-Dihydroxy-3-epi-vitamin D₃, a natural metabolite of 1a,25-dihydroxyvitamin D₃, is a potent suppressor of parathyroid hormone secretion. *J Cell Biochem*. 1999;73:106–113.
- 22. Siu-Caldera ML, Sekimoto H, Weiskopf A, et al. Production of 1a,25-dihydroxy-3-epi-vitamin D₃ in two rat osteosarcoma cell lines (UMR 106 and ROS 17/2.8): existence of the C-3 epimerization pathway in ROS 17/2.8 cells in which the C-24 oxidation pathway is not expressed. *Bone*. 1999;24:457–463.
- Bischof MG, Siu-Caldera M-L, Weiskopf A, et al. Differentiation-related pathways 1α,25-dihydroxycholecalciferol metabolism in human colon adenocarcinoma-derived caco-2 cells: production of 1α-25-dihydroxy-3-epi-cholecalciferol. *Exp Cell Res.* 1998;241:194–201.
- Siu-Caldera M-L, Sekimoto H, Brem AS, et al. Metabolism of la,25-dihydroxyvitamin D₃ into 1a,25-dihydroxy-3-epi-vitamin D₃ in rat osteosarcoma cells (UMR 106 and ROS 17/2.8) and rat vascular smooth muscle cells. *J Bone Min Res.* 1998;25(suppl):S426.
- 25. Kamao M, Tatematsu S, Hatakeyama S, et al. C-3 epimerization of vitamin D_3 metabolites and further metabolism of C-3 epimers:

25-hydroxyvitamin D_3 is metabolized to 3-epi-25-hydroxyvitamin D_3 and subsequently metabolized through C-1alpha or C-24 hydroxylation. *J Biol Chem.* 2004;279:15897–15907.

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- Kamao M, Tatematsu S, Sawada N, et al. Cell specificity and properties of the C-3 epimerization of vitamin D3 metabolites. *J Steroid Biochem Mol Biol*. 2004;89–90:39–42.
- Norman AW, Bouillon R, Farach-Carson MC, et al. Demonstration that 1 beta,25-dihydroxyvitamin D₃ is an antagonist of the nongenomic but not genomic biological responses and biological profile of the three A-ring diastereomers of 1 alpha,25-dihydroxyvitamin D₃. *J Biol Chem.* 1993;268:20022–20030.
- Molnár F, Sigüeiro R, Sato Y, et al. 1 a,25(OH)2-3-epi-vitamin D₃, a natural physiological metabolite of vitamin D₃: its synthesis, biological activity and crystal structure with its receptor. *PLoS One*. 2011;6:e18124.
- Rollins R. Vitamin D testing—what's the right answer? *Clin Lab News*. 2009;35(7):1.
- Fraser WD, Milan AM. Vitamin d assays: past and present debates, difficulties, and developments. *Calcif Tissue Int.* 2013;92:118–127.
- Couchman L, Benton CM, Moniz CF. Variability in the analysis of 25-hydroxyvitamin D by liquid chromatography-tandem mass spectrometry: the devil is in the detail. *Clin Chim Acta*. 2013;413: 1239–1243.
- Phinney KW, Bedner M, Tai SS, et al. Development and certification of a standard reference material for vitamin D metabolites in human serum. *Anal Chem.* 2012;84:956–962.
- 33. Singh RJ, Taylor RL, Satyanarayana R, et al. C-3 epimers can account for a significant proportion of total circulating 25-hydroxyvitamin D in infants, complicating accurate measurement and interpretation of vitamin D status. J Clin Endocrinol Metab. 2006;91:3055–3061.
- Stepman HC, Vanderroost A, Stöckl D, et al. Full-scan mass spectral evidence for 3-epi-25-hydroxyvitamin D in serum of infants and adults. *Clin Chem Lab Med.* 2011;49:253–256.
- 35. van den Ouwelanc JM, Beijers AM, van Daal H. Fast separation of 25-hydroxyvitamin D₃ from 3-epi-25-hydroxyvitamin D₃ in human serum by liquid chromatography-tandem mass spectrometry: variable prevalence of 3-epi-25-hydroxyvitamin D₃ in infants, children, and adults. *Clin Chem.* 2011;57:1618–1619.
- 36. Schleicher RL, Encisco SE, Chaudhary-Webb M, et al. Isotope dilution ultra performance liquid chromatography-tandem mass spectrometry method for simultaneous measurement of 25-hydroxyvitamin D2, 25-hydroxyvitamin D₃ and 3-epi-25-hydroxyvitamin D₃ in human serum. *Clin Chim Acta*. 2011;412:1594–1599.
- Strathmann FG, Sadilkova K, Laha, T, et al. 3-epi-25 hydroxyvitamin D concentrations are not correlated with age in a cohort of infants and adults. *Clin Chim Acta*. 2012;413:203–206.
- Lensmeyer G, Poquette M, Wiebe D, et al. The C-3 epimer of 25-hydroxyvitamin D(3) is present in adult serum. J Clin Endocrinol Metab. 2012;97:163–168.
- Baecher S, Leinenbach A, Wright JA, et al. Simultaneous quantification of four vitamin D metabolites in human serum using high performance liquid chromatography tandem mass spectrometry for vitamin D profiling. *Clin Biochem.* 2012;45:1491–1496.
- Bailey D, Veljkovic K, Yazdanpanah M, et al. Analytical measurement and clinical relevance of vitamin D(3) C3-epimer. *Clin Biochem.* 2013;46:190–196.

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