Potential use of biomarkers for the clinical evaluation of sarcoidosis

Amir Mousapasandi , ¹ Cristan Herbert, ¹ Paul Thomas ^{1,2}

¹School of Medical Sciences, Faculty of Medicine, University of New South Wales, Sydney, New South Wales, Australia ²Department of Respiratory Medicine, Prince of Wales' Hospital and Prince of Wales' Clinical School, University of New South Wales, Sydney, New South Wales, Australia

Correspondence to

Professor Paul Thomas, Department of Respiratory Medicine, Prince of Wales Hospital and Community Health Services, Sydney, NSW 2031, Australia; paul.thomas@unsw.edu.au

Accepted 29 December 2020 Published Online First 15 January 2021

ABSTRACT

Sarcoidosis is a systemic granulomatous disease of unknown etiology and pathogenesis with a heterogeneous clinical presentation. In the appropriate clinical and radiological context and with the exclusion of other diagnoses, the disease is characterized by the pathological presence of non-caseating epithelioid cell granulomas. Sarcoidosis is postulated to be a multifactorial disease caused by chronic antigenic stimulation. The immunopathogenesis of sarcoidosis encompasses a complex interaction between the host, genetic factors and postulated environmental and infectious triggers, which result in granuloma development. The exact pathogenesis of the disease has yet to be elucidated, but some of the inflammatory pathways that play a key role in disease progression and outcomes are becoming apparent, and these may form the logical basis for selecting potential biomarkers.

Biomarkers are biological molecules that are altered pathologically. To date, there exists no single reliable biomarker for the evaluation of sarcoidosis, either diagnostically or prognostically but new candidates are emerging. A diagnosis of sarcoidosis ideally requires a biopsy confirming non-caseating granulomas, but the likelihood of progression that requires intervention remains unpredictable. These challenging aspects could be potentially resolved by incorporating biomarkers into clinical practice for both diagnosis and monitoring disease activity. This review outlines the current knowledge on sarcoidosis with an emphasis on pulmonary sarcoidosis, and delineates the understanding surrounding the implication of biomarkers for the clinical evaluation of sarcoidosis.

BACKGROUND Enidemiology of sarco

Epidemiology of sarcoidosis

The incidence and prevalence of sarcoidosis is reported to be highest in Nordic countries (an incidence of 5–40 per 100 000 per year and a prevalence of 0.16%) and in African-Americans (incidence of 17.8–46 per 100 000 and a prevalence of 0.14%). The incidence is reported to be significantly lower in East Asia countries with Japan having an overall incidence of 1.01 per 100 000 and Korea having an incidence rate of 0.85 per 100 000.

The peak age for sarcoidosis onset ranges from 30 to 55 years. Gender plays a role in

manifestation of sarcoidosis, as males with sarcoidosis are diagnosed 3–10 years earlier than females who also have a higher prevalence. Mortality rates in sarcoidosis ranges from 1% to 8% depending on the type and location of the disease and other health factors. The disease are sarcoidosis ranges from 1% to 8% depending on the type and location of the disease and other health factors.

Etiology of sarcoidosis

The exact etiology of sarcoidosis remains unknown with no single genetic, infectious or environmental factor being identified to have a causal link to sarcoidosis.⁸

The current pathophysiological concept suggests a model in which sarcoidosis is caused by the combination of genetic polymorphisms creating a tendency to a specific immune response, associated with exposure to environmental or infectious agents. The sequence of events for the progression of sarcoidosis is depicted in figure 1. 10 11

Genetics

A genetic tendency in the development of sarcoidosis is demonstrated by familial clustering of sarcoidosis, the varying prevalence and subtype of presentation among different ethnicities and a higher occurrence in twin studies. African-Americans have a higher prevalence of sarcoidosis (3.8-fold to 4.0-fold greater risk than European Americans), a higher rate of extrathoracic involvement and more chronic and severe disease with a lower rate of remission. Twins studies indicate an 80-fold increased risk among monozygotic twins, and a 7-fold increase among dizygotic twins. ¹²⁻¹⁴

Case-control studies have identified that HLA alleles, responsible for the CD4+ T lymphocyte polypeptides in the HLA class I and II antigens, as being associated with specific disease subtypes and tendencies to sarcoidosis, for example, Lofgren's syndrome, a type of sarcoidosis with a good prognosis, being linked to the HLA-DR3 allele, in contrast to the HLA-DR15 allele which carries a worse prognosis. ¹⁵ These discoveries were further expanded by Genome Wide Association Studies and Case-Control Etiologic Study of Sarcoidosis (ACCESS) that also show the importance of genetic components in sarcoidosis. ¹¹ ¹⁷



© American Federation for Medical Research 2021. No commercial re-use. See rights and permissions. Published by BMJ.

To cite: Mousapasandi A, Herbert C, Thomas P. *J Investig Med* 2021;**69**:804–813.



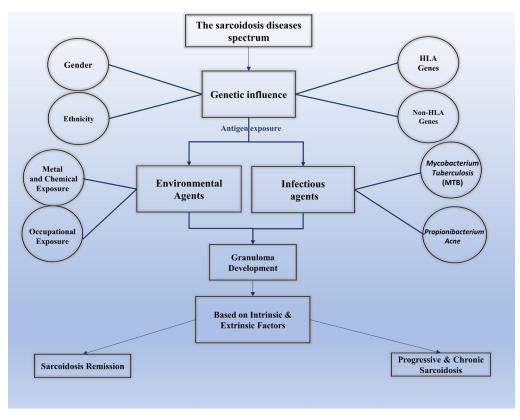


Figure 1 The sequence of events with the influence of different etiologies leading to granuloma formation.

Infectious agents

The radiological, clinical and immunological similarities between tuberculosis (TB) and sarcoidosis suggest the possible role of mycobacteria as an etiological agent. 18 The OR for the presence of mycobacterial DNA in tissue samples of patients with sarcoidosis compared with those from healthy subjects was found to be 9-fold to 19-fold and mycobacterial DNA was present in almost 30% of sarcoidosis biopsy specimens from sites such lymph nodes, lung, skin and others, 19-21 but to date, the culture of live organisms has been rare. Propionibacterium acnes has been isolated from culture in 78% of sarcoid lesions, which suggest a role for this organism in sarcoidosis; however, P. acnes was also found in 20% of non-sarcoid lymph nodes. 22 Animal models have also indicated that P. acnes can induce antigen-driven granulomatous inflammation.²³ Diseases with similar pathological and immunological features that resemble sarcoidosis, such as hypersensitivity pneumonitis and chronic beryllium disease, indicate that it is unclear whether sarcoidosis has an infectious etiology or not and more studies on the subject are required to investigate the exact role of these putative infectious agents.^{24–26}

Environmental agents

Exposure to metals and minerals such as beryllium, chromium, aluminum, titanium, zirconium, talc and nickel has been shown to induce sarcoid-like granulomas.²⁷ Epidemiological studies have identified positive associations between occupations such as metal-working, fire-fighting and the handling of building supplies and sarcoidosis.¹¹ ²⁸ ²⁹

Tobacco smoking decreases the risk of sarcoidosis, possibly because smoking deactivates M2 alveolar macrophages and the macrophage is thought to be pivotal in the pathogenesis of the disease. The ACCESS studies have identified several environmental exposures associated with an increased risk of sarcoidosis including agricultural materials, pesticides, insecticides and microbial aerosols (mouldy and musty odours). Although there are many findings implicating environmental agents as risk factors, the current evidence does not strongly favor a single environmental or occupational exposure, and may implicate a range of precipitants.

Pathogenesis of sarcoidosis

The histopathological appearance of sarcoidosis is that of non-caseating granulomas formed as the result of aberrant cell-mediated immune responses to unknown antigens. Sarcoid granulomas are characterized by a central core of giant cells, epithelioid cells and helper T cells (Th). 16 32 This central area is surrounded by monocytes, mast cells, CD8+ and CD4+ T lymphocytes, B lymphocytes and fibroblasts, which in turn are surrounded by lamellar rings of hyaline collagen (figure 1). The proportions of lymphocytic infiltrate and fibrosis surrounding the central core vary depending on the patient and disease duration. Additional histopathological elements of sarcoid granulomas that may be present include Schaumann bodies, birefringent crystalline particles and asteroid bodies, which are biomolecules and mineral components incorporated into the granulomas.33

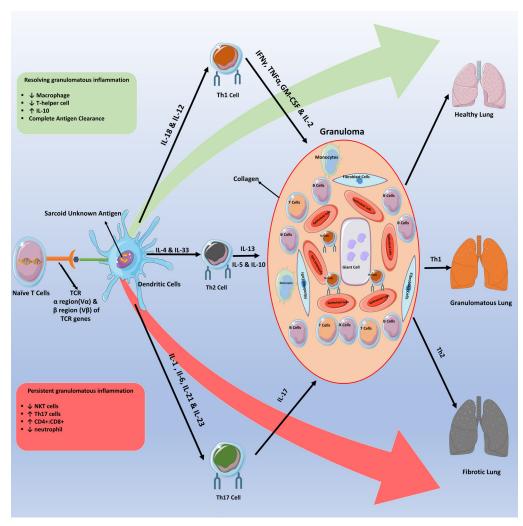


Figure 2 Overview of the postulated immunopathogenesis of sarcoidosis. The immunopathogenesis of sarcoidosis showing three different outcomes of the disease and the dominant immunological markers involved in each. GM-CSF, granulocyte-macrophage colony-stimulating factor; IL, interleukin; IFN- γ , interferon gamma; NKT, natural killer cells; TCR, T-cell receptors; TNF- α , tumor necrosis factor alpha.

As a result of antigen-presentation by dendritic cells and subsequent T cell activation, monocytes are recruited to the site of inflammation. In the case of incomplete antigenic clearance, the monocytes build up and eventually mature into epithelioid cells. Following the cell-mediated immune responses, T lymphocytes are also recruited and infiltrate the tissue, and a granuloma develops (figure 2). A sarcoid granuloma is thought to be a dynamic structure recruiting newly formed monocytes, which gradually penetrate the core where they mature into epithelioid cells. ³⁴ ³⁵

Th1 cells

Interleukin-12 and interleukin-18 (IL-12 and IL-18 as cytokines) are activated and released by dendritic cells to promote Th1 proliferation and interferon gamma (IFN-γ) production.³⁶ Subsequently, IL-18 amplifies the expression of tumour necrosis factor alpha (TNF-α), IL-2 and granulocyte-macrophage colony-stimulating factor (GM-CSF) that are produced from activated dendritic cells which in turn enhance the CD4+T cells and formation of granulomatous inflammation in a synergistic manner.^{37 38}

IFN- γ promotes the production of T-bet and the chemokine receptor CXCR3 that result in a positive feedback-loop which further increases IFN- γ through Th1 mechanisms. Increased levels of IFN- γ from the feedback-loop decreases the expression of Th2 cytokine transcription and the production of Th2 cytokines such as IL-4 and IL-13. ^{39 40}

Despite the dominant role of Th1 cells in sarcoidosis, recent studies show that Th17 cells also play a role. Th17 cells produce IL-17A which is vital for formation of mature granulomas. Additionally, Th17 cells appear to be an important Th cell subtype in bronchoalveolar lavage fluid (BALF) of patients with sarcoidosis and have elevated levels of IFN-γ. 41-44

Th2 cells

Fibrosis in sarcoidosis is thought to be caused by the shift in the immune response from Th1 to Th2 dominance.⁴⁵ IL-13, a Th2 cytokine, stimulates transforming growth factor beta (TGF-β) activity which induce the transformation of fibroblasts into myofibroblasts. Fibrosis in sarcoidosis may also be related to the role of elevated chemokine ligand

2 (CCL2, a Th2 chemokine) as it prevents fibroblast cell death through IL-6/STAT3 signaling. 46 47 A summary of the contribution of Th1, Th17 and Th2 cells to the pathogenesis of sarcoidosis is depicted in figure 2.

Clinical diagnosis

Currently, most of the diagnostic and monitoring tests available require an invasive biopsy. Hence, development of a less invasive test for sarcoidosis is a goal of ongoing research. To date, there exist no 'gold standard' diagnostic criteria. However, a presumptive diagnosis of sarcoidosis can be made following radiological evidence of sarcoidosis such as bilateral symmetrical hilar lymphadenopathy, clinical evidence of sarcoidosis such as erythema nodosum and pulmonary involvement or histological finding of non-caseating granuloma. The latest American Thoracic Society clinical practice guidelines do not make a definitive statement in relation to biopsy. Endobronchial ultrasound and biopsy has a high yield, but depends on the presence of proximal mediastinal or hilar lymphadenopathy.

The presentation of sarcoidosis may be an incidental asymptomatic finding on imaging, such as intrathoracic lymphadenopathy on chest radiograph in 87% of cases or by clinical presentation with cough or breathlessness in symptomatic individuals. Pulmonary disease is present in up to 97% of cases, together with cutaneous, hepatosplenomegaly, central nervous system (CNS), ocular and cardiac sarcoidosis.^{3 50}

Spontaneous remission occurs in most cases of sarcoidosis, but 25%–40% of patients will have a chronic and protracted course and it can be the cause of death in approximately 5% of cases.⁵¹

BIOMARKERS FOR CLINICAL EVALUATION

Biomarkers can be considered as indicators of normal or pathological processes, and the use of potential biomarkers to provide a diagnosis and to monitor disease activity remain the focus of ongoing research. Several approaches have been adopted to detect novel biomarkers in BALF, exhaled breath condensate (EBC) and serum using proteomic analysis, ELISA of specific candidate meditators and genome-based approaches. ^{52–54} Many of these reports require to be conducted in large-scale studies for better external validity and to be able to distinguish the different phases of the disease such as radiological staging, lung function and extrathoracic involvement. ^{52–55}

Furthermore, studies identifying novel biomarkers may guide approaches to expand our understanding of this puzzling disease. Despite many potential biomarkers being recognized, a lack of sensitivity and specificity have hampered clinical usage, therefore necessitating further research in this area. ⁵⁶

Serum

The traditional approach to investigate the pathogenesis of a disease is by examining the serum biomarkers. Initially, it was believed that analyzing serological biomarkers would be beneficial in sarcoidosis as sarcoidosis is a multiorgan disease and these might better represent the underlying systemic inflammation, but local inflammation can be assessed, for example, in the breath.^{57 58} However, despite

many such biomarkers being discovered to be associated with sarcoidosis, none has been shown to have the adequate accuracy to be used for routine diagnosis, with the possible exception of ACE. Some serum biomarker levels may be altered in single organ disease, but not in others, for example, in pulmonary sarcoidosis the systemic depletion of peripheral lymphocytes (peripheral anergy) may lead to serum biomarkers not being representative of the disease state in pulmonary sarcoidosis, whereas in, for example, solitary sarcoid uveitis, systemic markers may not be altered at all.⁵⁹

Potential serum biomarkers in patients with sarcoidosis have included angiotensin converting enzyme (ACE), serum amyloid A, cytokines, chemokines, microRNAs (miRNAs), chitotriosidase and lysosomes as summarized in table 1.60-77 Serum chitotriosidase was shown to have relatively accurate diagnostic ability with very high sensitivity and specificity, however, another study with different cutoff values indicated substantially lower diagnostic values (sensitivity and specificity of 88.6% and 92.8% for cutoff of 48.8 nmol/hour/mL and 82.5% and 70% for cut-off of 100 nmol/hour/mL, respectively).62 78 Further research should be conducted to correctly delineate the cut-off value that provides optimal diagnostic accuracy, and with more sample size and clinical context, however, it is possible to have different cut-offs for various types of patients or clinical backgrounds.

To date, only ACE been adopted for clinical usage due to the low sensitivity/specificity and lack of reproducibility of other potential novel biomarkers, although chitotriosidase may be useful for disease monitoring, progression and response to treatment. A study assessing the ACE levels of 3277 patients with sarcoidosis concluded that despite the common practice, ACE should not be used in clinical practice for the diagnosis of sarcoidosis.⁷⁹

Serum biomarkers could be valuable in distinguishing between granulomatous diseases such as TB and sarcoidosis. Serum leptin and intercellular adhesion molecule 1 levels were shown to be significantly elevated in patients with sarcoidosis rather than in TB.80 Another serum biomarker, adiponectin, which is an anti-inflammatory protein was differentially expressed in patients with sarcoidosis compared with healthy individuals.⁸¹ Soluble CD163 was also significantly elevated in patients with sarcoidosis and its level associated with serum ACE and soluble IL-2 receptor levels, an important marker of macrophage activity involved in sarcoidosis, hence CD163 could provide insight on important aspects of disease activity.⁸² Biomarkers could be used to identify the organ involved in sarcoidosis. Serum ACE and soluble IL-2R levels were significantly lower in isolated cardiac sarcoidosis than systemic sarcoidosis, whereas B-type natriuretic peptide (BNP) was elevated in cardiac sarcoidosis, thus BNP might be a useful marker for detecting cardiac involvement in conjunction with other clinical tests.⁸³ Advanced imaging for cardiac sarcoidosis such as cardiac magnetic resonance and ^{18F}FDG PET have proven valuable in establishing clinical assessment of the disease, and these techniques coupled with specific organ biomarkers could provide a possible diagnostic approach when biopsy is difficult to perform in extrathoracic organ involvement.84 Similarly, attempts have been made to identify serum neurosarcoidosis biomarkers, and serum S100B

Table 1 Serum biomarkers in patients with sarcoidosis Change in patients with Serum hiomarkers sarcoidosis Clinical significance ACE (produced by epithelioid cells derived from \uparrow serum ACE $\rightarrow \uparrow$ in granuloma formation, extrathoracic involvement and disease activity6 activated macrophages) Sensitivity ranges from 41% to 100% and specificity ranges from 83% to 99% for diagnosis of sarcoidosis⁶⁷ Soluble interleukin (IL)-2 receptor ↑ sIL-2R → ↑ extrathoracic organs involvement and disease severity 66 75 (sIL-2R) (marker for T-cell activation) The sensitivity and specificity of ACE are 62% and 76%, respectively 64 Soluble CD163 (sCD163) ↑ sCD163 \rightarrow ↑ serum ACE and soluble ↑ sIL-2 ↑ sCD163 correlates with disease activity ↑ (An inverse relationship with ↑ CXCL-9 and CXCL-10 \rightarrow More chronic and severe form of sarcoidosis^{73 74} Chemokines (induced by interferon (IFN)-γ) pulmonary function) Chitotriosidase ↑ CTO correlates with disease severity⁶² (CTO) (serum marker of macrophage activation) The sensitivity and specificity of CTO are 89% and 93%, respectively with a cut-off of 48.8 nmol/hour/mL62 ↑ SAA in patients with sarcoidosis 63 Serum amyloid A (an acute phase protein stimulated by IL-1 and IL-6) ↑↑ SAA in active sarcoidosis⁷⁷ Serum lysozyme Serum lysozyme correlates with radiographic stage⁷⁰ (bacteriolytic enzyme present in macrophages) MicroRNAs (miRNAs) (regulation of gene expression) ↑ hsa-miRNA-128-3p, hsa-miRNA-22-5p, hsa-miRNA-30e-3p, hsamiRNA-4306, hsa-miRNA-92a-1-5p, hsa-miRNA-150-3p, hsa-miRNA-6729-5p and hsa-miRNA-342-5p \rightarrow Potential initial diagnostic biomarker⁶¹ sensitivity of 74.8% and positive predicted value of 88.24%61 \uparrow IL-18 and IL-12 \rightarrow \uparrow IFN- γ correlates with sarcoidosis activity $^{65~68~71~72}$

which is a calcium-binding protein was found to be elevated and correlated with CNS injury in neurosarcoidosis. 82

1

Neurosarcoidosis and cardiac sarcoidosis remain difficult to diagnose and remain based on a combined clinical picture with MRI/PET and other features such as CSF ACE and oligoclonal bands, or the presence of more accessible disease to biopsy.

Bronchoalveolar lavage fluid

(synthesized by CD4+ T cells and macrophages)

Interleukins (IL)

Since pulmonary disease is common, sampling the lung could provide valuable insight about sarcoidosis activity and clinical evaluation. Changes in the inflammatory profile in BALF or breath could reflect the activity of components of sarcoidosis-related inflammation, that is, macrophages and lymphocytes. 86 Sarcoidosis is characterized by an increase in BALF lymphocytes and the CD4/CD8 ratio (1.7±1 in healthy individuals vs 9.3±5.0 in symptomatic individuals). 86 A meta-analysis demonstrated that the BALF CD4/ CD8 ratio provides a sensitivity of 70% and specificity of 83% for sarcoidosis, a better accuracy than ACE, a marker commonly used in clinical practice. No clear cut-off value was found as the values ranged from 2 to 4 between studies, therefore it is crucial to find a value to provide optimal diagnostic accuracy. The value, CD4/CD8 ratio ≥3.5, was useful for patients who presented with typical clinical and radiological manifestation of the disease, but, the ratio is not selective enough to be employed on its own and must be integrated with other established diagnostic methods.⁸⁷

Furthermore, matrix metalloproteinase 12 (MMP12), an elastase enzyme produced by macrophages is known to have elevated gene and protein expression in BALF of patients with sarcoidosis and the levels were correlated with disease severity.⁸⁸ The altered MMP12 expression in BALF resulted in animal studies being conducted, which

demonstrated that MMP12 knockout mice had a significant reduction in granuloma formation and reduced expression of IFN-y, an important mediator in sarcoidosis pathogenicity. These findings suggest a critical role for MMP12 in the chronicity of granulomatous inflammation. 89 Another BALF biomarker is the T-cell subset CD4+ $V\alpha 2.3+$, which is associated with a better prognosis and potentially may be used as a surrogate prognostic marker. 90

As in serum, a range of chemokines, cytokines, ILs and lymphocytes are altered in the BALF of patients with sarcoidosis, as summarized in table 2. For instance, patients with Löfgren's syndrome, who have a better prognosis, were established to have lower IFN-γ and TNF-α messenger RNA (mRNA) levels in their BALF, pro-inflammatory mediators involved in sarcoidosis pathology, than HLA subtypes with worse prognosis.⁹¹

Exhaled breath condensate

An alternative method to assess the BALF is by collecting EBC which is a non-invasive method of collecting exhaled breath containing the airway lining fluid (ALF) and soluble exhaled gases. The ALF can be analyzed by collecting BALF, however collection requires bronchoscopy and therefore is not a feasible approach for routine monitoring. 92 Moreover, bronchoscopy is an invasive technique that has complications such as pneumothorax (0%–4%), desaturation (0.7%– 76.3%), bleeding (2.5%–89.9%), arrhythmia (8%–25.7%) and patient discomfort (55.4%-96.3%), making it impractical as a repeated routine test and would likely have low acceptance as a frequent test. 93

EBC samples the ALF, and therefore can assess airway inflammation and disease activity 94 95 with potential EBC biomarkers linked to the underlying pathophysiology of respiratory conditions, including asthma, cystic fibrosis,

Table 2 BALF biomarkers in patients with sarcoidosis		
BALF biomarkers	Change in sarcoidosis	Significance
MicroRNAs (miRNAs)	↑ and ↓	↑ miRNA-146a and miRNA-150 (extracellular) and miR-21 (cellular) in CXR-II compared with CXR-I (chest-X-ray-II stage more advanced than I) ¹⁰⁵ ↑ miRNA-04, miRNA-146a, miRNA-150 and miRNA-222 and ↓ miR-202 and miR-204 in sarcoidosis disease ¹³³ ↑ miRNA-27b, miRNA-192 and miRNA-221(involved in angiogenesis) in acute sarcoidosis
Cytokines	↑	↑ IL-18 correlates with sarcoidosis activity ^{68 135} ↑ IL-33 correlates with diffusion lung capacity for carbon monoxide ¹³⁶ ↑INF- γ and TNF- α (Th1 cytokines) ⁹¹ ↑ IL-2, IL-4, IL-10, IL-12 and IL-13 (Th2 cytokines) ¹³⁷
Lymphocytes	↑	↑ Lymphocyte T CD4+/CD8+ correlates with active disease state and ↑ Lymphocyte T IL-17+/CD4+ correlates with active disease state ⁴³ CD4+/CD8+ cut-off value not clear
Chemokines	↑	↑ CCL2 and CCL5 (chemokine ligands recruiting monocytes) in all stages of disease ¹⁰¹
Matrix metalloproteinase 12 (MMP12)	↑	\uparrow MMP12 $\to \uparrow$ granuloma formation and \uparrow IFN- γ expression \uparrow MMP12 correlates with disease severity

IFN, interferon; IL, interleukin; TNF, tumor necrosis factor.

chronic obstructive pulmonary disease and sarcoidosis. 72 $^{96-99}$

The concentration of total protein and some inflammatory modulators are significantly elevated in EBC from sarcoidosis compared with healthy subjects and this elevation is in agreement with studies of BALF. A subanalysis of these proteins and inflammatory markers is required to specifically identify the important cytokines and signaling proteins involved in sarcoidosis and are differentially expressed. $^{100-103}$ Original work from our laboratory has demonstrated that EBC TGF- $\beta 1$, an inflammatory mediator involved in sarcoidosis, and neopterin are elevated in patients with sarcoidosis compared with healthy individuals. 102

Another advantage of evaluating EBC biomarkers is that the technique is totally non-invasive. Such a measurement has potential application in serial monitoring of disease activity which lends itself to being able to indicate regression or relapse. Potential biomarkers in BALF and EBC such as TNF-α, miRNAs, cytokines, IFN-γ and extracellular vesicles (EV) (exosomes) have been found to differ between patients with sarcoidosis and healthy controls and hence identified as possible diagnostic and prognostic tools which need to be tested in prospective studies. 60 100 104-107 In order to further develop the EBC technique in clinical care, much work has been focused on standardisation of EBC collection for use in clinical practice.

A summary of key studies which demonstrate the BALF biomarkers that are differentially expressed in patients with sarcoidosis compared with healthy controls is demonstrated in table 2 but more work is required to better characterize EBC findings in sarcoidosis and establish those that are most reliable.

Limitation of biomarkers

The use of biomarkers for sarcoidosis may be challenging for several reasons. First, the criteria for the diagnosis for the diagnosis of sarcoidosis rely on the clinical presentation, biopsy result with non-caseating granulomata and the exclusion of other diagnoses, which can affect the certainty of a diagnosis when assessing the accuracy of a newly developed diagnostic biomarker. ¹⁰⁸ Furthermore, sarcoidosis

is a systemic disease in which multiple organs may be affected to a varying extent. Therefore, unidimensional biomarkers are unlikely to encapsulate the whole spectrum of the disease. Techniques to overcome this problem are to use a combination of several biomarkers or to study those related to specific organ involvement. ¹⁰⁹ In addition, it is likely that some biomarkers will reflect active macrophage and lymphocytic inflammation, while other may reflect the healing process, and yet others represent a pathway to ongoing fibrosis. ⁴³

Despite some research, no novel biomarkers have been adopted into clinical practice due to their relatively low accuracy, sensitivity and specificity. ⁶⁰ Therefore, biomarkers remain an area that requires more study.

DISCOVERY AND VALIDATION OF NOVEL BIOMARKERS

While many advances have been made in understanding the pathogenesis and the identification of biomarkers that are plausible indicators of sarcoidosis, novel aspects of the basic mechanisms are becoming apparent as being involved in disease progression and resolution and may offer opportunities for novel biomarkers. Some of these novel biomarkers that have shown initial success and require more research include miRNAs and exosomes.

MicroRNAs

miRNAs are a class of regulatory molecules suggested to be as prospective biomarkers with the possible involvement in sarcoidosis pathophysiology. $^{104\,110}$ miRNAs are non-coding, single-stranded RNAs composed of 18–25 nucleotides, and the importance of miRNA altered regulation in lung disease is becoming increasingly evident. 111 miRNAs likely play a role in both inflammation and granuloma formation, hence a significant component of sarcoidosis pathophysiology. 112 miRNAs are able to change the function of various inflammatory and apoptosis signaling pathways by inhibiting the post-transcriptional gene expression via mRNA or altering protein translation. $^{111\,113}$ The inflammatory regulation related to sarcoidosis includes modulation of T-cell differentiation, IFN- γ expression, Th1 cells and IL-2R. 114

Review

Multiple studies demonstrate the relationship between dysregulation of several miRNAs and sarcoidosis. 115 116 As highlighted in BALF and serum, many miRNAs have altered expression in sarcoidosis, and some are associated with disease severity. It is unclear whether these miRNAs would be elevated in sarcoidosis EBC samples, complicated by the fact that extracellular miRNAs are mostly encapsulated inside EV. 117

Exosomes

Exosomes are small, 30–120 nm in diameter, cell-derived EV and are secreted from most cell types. ¹¹⁷ Exosomes mainly serve as a means of protection to transfer cell surface molecules, proteins, miRNAs and DNAs to specific location for intercellular communication and to protect them from enzymatic breakdown. ¹¹⁸

Exosomes are biologically active EV that are tightly regulated thorough various mediators and pathways. miRNAs are mostly encapsulated within exosomes and therefore exosome isolation improves the sensitivity of miRNA expression and identification, ¹¹⁹ although not all studies have confirmed this view with miRNAs being also present outside exosomes and bound to proteins such as Argonaute 2. ¹²⁰ 121

The number and the contents of exosomes in body fluids increase in cancer and sarcoidosis, however exosomal function in sarcoidosis pathophysiology remains poorly understood. 107 122 Sarcoidosis BALF exosomal miRNA and cytokines were elevated, for example, miRNA-146a. 107 123 BALF exosomes from patients with sarcoidosis induced higher levels of IFN-γ and IL-13 production by epithelial cells therefore illustrating an association with the underlying immunopathogenesis of sarcoidosis. 107 We have shown that EBC exosomes are able to be isolated and exosome-induced production of TNF-α was greater in monocytes from patients than from controls and miRNAs and mRNA involved in sarcoidosis pathology was expressed higher in serum exosomes of patients.¹²⁴ A number of studies have investigated miRNA expression in pulmonary sarcoidosis. 114 Several investigated miRNA expression in peripheral blood, blood cells bronchoalveolar cells, exosomal/EV miRNA in BALF but none studied exosomal miRNA expression in EBC. 61 115 125-131 Thus, there is a need for more studies to be conducted with larger numbers of subjects, different sample sources such as EBC and to target known mediators of sarcoidosis.

Moreover, the exosomal miRNAs changes were not limited to pulmonary sarcoidosis as a study indicated several exososomal miRNAs levels were higher in patients with cardiac sarcoidosis, hence further illustrating the potential of miRNAs as biomarkers and the possibility of using specific biomarkers for detecting vital organ involvement.¹³²

CONCLUSION AND FUTURE RESEARCH

Evaluating exosomal and non-exosomal miRNAs and cytokine expression in EBC samples has remained an unexplored area in the literature and thus requires further investigation. Furthermore, our preliminary data indicate that exosomes and miRNAs regulating key sarcoidosis cytokines that are differentially expressed in EBC, hence more research is required to identify and validate sarcoidosis biomarkers that have the advantage of being non-invasive, sensitive and amenable to repeated sampling. 124 Many recent studies have demonstrated a variety of mediators involved in the immunopathogenesis of sarcoidosis and the levels were shown to be differentially expressed in sarcoidosis, indicating that they may be potential biomarkers. However, uncertainty remains as to which aspect of clinical assessment, diagnosis, prognosis or disease activity, these biomarkers could be applied. The pattern in which biomarkers were expressed in organ-specific sarcoidosis is not always consistent with other forms of the diseases, hence more focused studies are mandated to better characterize the nature of biomarkers in different disease forms. Likewise, long-term studies with a baseline profile of the potential markers would be helpful in indicating prognosis, but the analysis will be complex with many variables including gender, ethnicity, MHC/genomic and other variables requiring assessment, and the results will need to be validated carefully.

Twitter Amir Mousapasandi @AmirMP_

Acknowledgements The original figure (figure 2) was created using Servier Medical Art Templates, which are licensed under a Creative Common Attribution 3.0 Unported License: https://smart.servier.com.

Contributors AM planned and wrote the initial draft of the manuscript; CH and PT edited the manuscript for submission.

Funding The authors have not declared a specific grant for this research from any funding agency in the public, commercial or not-for-profit sectors.

Competing interests None declared.

Patient consent for publication Not required.

Provenance and peer review Not commissioned; externally peer reviewed.

ORCID iD

Amir Mousapasandi http://orcid.org/0000-0003-0636-2419

REFERENCES

- 1 Arkema EV, Grunewald J, Kullberg S, et al. Sarcoidosis incidence and prevalence: a nationwide register-based assessment in Sweden. Eur Respir J 2016:48:1690–9.
- 2 Baughman RP, Field S, Costabel U, et al. Sarcoidosis in America. analysis based on health care use. Ann Am Thorac Soc 2016;13:1244–52.
- 3 Ungprasert P, Carmona EM, Utz JP, et al. Epidemiology of sarcoidosis 1946-2013: a population-based study. Mayo Clin Proc 2016;91:183–8.
- 4 Yoon H-Y, Kim HM, Kim Y-J, et al. Prevalence and incidence of sarcoidosis in Korea: a nationwide population-based study. *Respir Res* 2018;19:158.
- 5 Morimoto T, Azuma A, Abe S, et al. Epidemiology of sarcoidosis in Japan. Eur Respir J 2008;31:372–9.
- 6 Arkema EV, Cozier YC. Epidemiology of sarcoidosis: current findings and future directions. Ther Adv Chronic Dis 2018;9:227–40.
- 7 Gerke AK. Morbidity and mortality in sarcoidosis. Curr Opin Pulm Med 2014;20:472–8.
- 8 Ramachandraiah V, Aronow W, Chandy D. Pulmonary sarcoidosis: an update. Postgrad Med 2017;129:149–58.
- 9 Dubrey S, Shah S, Hardman T, et al. Sarcoidosis: the links between epidemiology and aetiology. Postgrad Med J 2014;90:582–9.
- 10 Semenzato G. Access: a case control etiologic study of sarcoidosis. Sarcoidosis Vasc Diffuse Lung Dis 2005;22:83–6.
- 11 Barnard J, Rose C, Newman L, et al. Job and industry classifications associated with sarcoidosis in a case-control etiologic study of sarcoidosis (access). J Occup Environ Med 2005;47:226–34.
- 12 Saidha S, Sotirchos ES, Eckstein C. Etiology of sarcoidosis: does infection play a role? Yale J Biol Med 2012;85:133–41.
- 13 Nessrine A, Zahra AF, Taoufik H. Musculoskeletal involvement in sarcoidosis. J Bras Pneumol 2014;40:175–82.
- 14 Sverrild A, Backer V, Kyvik KO, et al. Heredity in sarcoidosis: a registry-based twin study. *Thorax* 2008;63:894–6.
- 15 Grunewald J, Grutters JC, Arkema EV, et al. Sarcoidosis. Nat Rev Dis Primers 2019:5:45.

- 16 Valeyre D, Prasse A, Nunes H, et al. Sarcoidosis. The Lancet 2014;383:1155–67.
- 17 Adrianto I, Lin CP, Hale JJ, et al. Genome-Wide association study of African and European Americans implicates multiple shared and ethnic specific loci in sarcoidosis susceptibility. PLoS One 2012;7:e43907.
- 18 Drake WP, Pei Z, Pride DT, et al. Molecular analysis of sarcoidosis tissues for Mycobacterium species DNA. Emerg Infect Dis 2002;8:1334–41.
- 19 Dubaniewicz A, Trzonkowski P, Dubaniewicz-Wybieralska M, et al. Mycobacterial heat shock protein-induced blood T lymphocytes subsets and cytokine pattern: comparison of sarcoidosis with tuberculosis and healthy controls. Respirology 2007;12:346–54.
- 20 Gupta D, Agarwal R, Aggarwal AN, et al. Molecular evidence for the role of mycobacteria in sarcoidosis: a meta-analysis. Eur Respir J 2007;30:508–16.
- 21 Svendsen CB, Milman N, Rasmussen EM, et al. The continuing search for Mycobacterium tuberculosis involvement in sarcoidosis: a study on archival biopsy specimens. Clin Respir J 2011;5:99–104.
- 22 Asakawa N, Uchida K, Sakakibara M, et al. Immunohistochemical identification of Propionibacterium acnes in granuloma and inflammatory cells of myocardial tissues obtained from cardiac sarcoidosis patients. PLoS One 2017;12:e0179980.
- 23 Mukherjee S. Is *Propionibacterium acnes* the cause of sarcoidosis? *Thorax* 2004;59:942.
- 24 Celada LJ, Hawkins C, Drake WP. The etiologic role of infectious antigens in sarcoidosis pathogenesis. *Clin Chest Med* 2015;36:561–8.
- 25 DiGiulio DB, Romero R, Amogan HP, et al. Microbial prevalence, diversity and abundance in amniotic fluid during preterm labor: a molecular and culturebased investigation. PLoS One 2008;3:e3056.
- 26 Hanak V, Kalra S, Aksamit TR, et al. Hot tub lung: presenting features and clinical course of 21 patients. Respir Med 2006;100:610–5.
- 27 Newman KL, Newman LS. Occupational causes of sarcoidosis. Curr Opin Allergy Clin Immunol 2012;12:145–50.
- 28 Kucera GP, Rybicki BA, Kirkey KL, et al. Occupational risk factors for sarcoidosis in African-American siblings. Chest 2003;123:1527–35.
- 29 Prezant DJ, Dhala A, Goldstein A, et al. The incidence, prevalence, and severity of sarcoidosis in New York City firefighters. Chest 1999;116:1183–93.
- 30 Almatroodi SA, McDonald CF, Darby IA, et al. Characterization of M1/M2 tumour-associated macrophages (TAMs) and Th1/Th2 cytokine profiles in patients with NSCLC. Cancer Microenviron 2016;9:1–11.
- 31 Mroz MM, Maier LA, Strand M, et al. Beryllium lymphocyte proliferation test surveillance identifies clinically significant beryllium disease. Am J Ind Med 2009;52:762–73.
- 32 Rossi G, Cavazza A, Colby TV. Pathology of sarcoidosis. Clin Rev Allergy Immunol 2015;49:36–44.
- 33 Ahmadzai H, Huang S, Steinfort C, et al. Sarcoidosis: a state of the art review from the thoracic Society of Australia and New Zealand. Med J Aust 2018;208:499–504.
- 34 Mitchell DN, Scadding JG, Heard BE, et al. Sarcoidosis: histopathological definition and clinical diagnosis. J Clin Pathol 1977;30:395–408.
- 35 Soler P, Basset F, Bernaudin JF. Morphology and distribution of the cells of a sarcoid granuloma: ultrastructural study of serial sections. *Ann N Y Acad Sci* 1976;278:147–60.
- 36 Moller DR, Forman JD, Liu MC, et al. Enhanced expression of IL-12 associated with Th1 cytokine profiles in active pulmonary sarcoidosis. J Immunol 1996:156:4952.
- 37 Eubank TD, Roberts R, Galloway M, et al. GM-CSF induces expression of soluble VEGF receptor-1 from human monocytes and inhibits angiogenesis in mice. Immunity 2004;21:831–42.
- 38 Puren AJ, Fantuzzi G, Gu Y, et al. Interleukin-18 (IFNgamma-inducing factor) induces IL-8 and IL-1beta via TNFalpha production from non-CD14+ human blood mononuclear cells. J Clin Invest 1998;101:711–21.
- 39 Amsen D, Spilianakis CG, Flavell RA. How are T(H)1 and T(H)2 effector cells made? Curr Opin Immunol 2009;21:153–60.
- 40 Zissel G, Müller-Quernheim J. Cellular players in the immunopathogenesis of sarcoidosis. *Clin Chest Med* 2015;36:549–60.
- 41 Ramstein J, Broos CE, Simpson LJ, et al. IFN-γ-Producing T-helper 17.1 cells are increased in sarcoidosis and are more prevalent than T-helper type 1 cells. Am J Respir Crit Care Med 2016;193:1281–91.
- 42 Urbankowski T, Hoser G, Domagała-Kulawik J. Th1/Th2/Th17-related cytokines in the bronchoalveolar lavage fluid of patients with sarcoidosis: association with smoking. *Pol Arch Med Wewn* 2012;122:320–5.
- 43 Loke WSJ, Herbert C, Thomas PS. Sarcoidosis: immunopathogenesis and immunological markers. *Int J Chronic Dis* 2013;2013:1–13.
- 44 Arger NK, Machiraju S, Allen IE, et al. T-Bet expression in peripheral Th17.0 cells is associated with pulmonary function changes in sarcoidosis. Front Immunol 2020;11:1129.

- 45 Broos CE, Hendriks RW, Kool M. T-Cell immunology in sarcoidosis: disruption of a delicate balance between helper and regulatory T-cells. *Curr Opin Pulm Med* 2016:22:476–83.
- 46 Liu X, Das AM, Seideman J, et al. The CC chemokine ligand 2 (CCL2) mediates fibroblast survival through IL-6. Am J Respir Cell Mol Biol 2007;37:121–8.
- 47 Xaubet A, Marin-Arguedas A, Lario S, *et al*. Transforming growth factor-beta1 gene polymorphisms are associated with disease progression in idiopathic pulmonary fibrosis. *Am J Respir Crit Care Med* 2003;168:431–5.
- 48 Soto-Gomez N, Peters JI, Nambiar AM. Diagnosis and management of sarcoidosis. Am Fam Physician 2016;93:840–8.
- 49 Crouser ED, Maier LA, Wilson KC, et al. Diagnosis and detection of sarcoidosis. An official American thoracic Society clinical practice guideline. Am J Respir Crit Care Med 2020;201:e26–51.
- 50 Rao DA, Dellaripa PF. Extrapulmonary manifestations of sarcoidosis. Rheum Dis Clin North Am 2013;39:277–97.
- 51 Gerke AK, Judson MA, Cozier YC, et al. Disease burden and variability in sarcoidosis. *Ann Am Thorac Soc* 2017;14:S421–8.
- 52 Bonham CA. Biomarkers in sarcoidosis: can microRNAs fill the gap? Am J Respir Cell Mol Biol 2018;58:1–2.
- 53 Belhomme N, Jouneau S, Bouzillé G, et al. Role of serum immunoglobulins for predicting sarcoidosis outcome: a cohort study. PLoS One 2018;13:e0193122.
- 54 Casanova N, Zhou T, Knox KS, et al. Identifying novel biomarkers in sarcoidosis using genome-based approaches. Clin Chest Med 2015;36:621–30.
- 55 Bergantini L, Bianchi F, Cameli P, et al. Prognostic biomarkers of sarcoidosis: a comparative study of serum chitotriosidase, ACE, lysozyme, and KL-6. Dis Markers 2019;2019:1–7.
- 56 Ahmadzai H, Loke WSJ, Huang S, et al. Biomarkers in sarcoidosis: a review. Curr Biomark Find 2014;4:93–106.
- 57 Ahmadzai H, Wakefield D, Thomas PS. The potential of the immunological markers of sarcoidosis in exhaled breath and peripheral blood as future diagnostic and monitoring techniques. *Inflammopharmacology* 2011:19:55–68
- 58 Sng JJ, Prazakova S, Thomas PS, et al. MMP-8, MMP-9 and neutrophil elastase in peripheral blood and exhaled breath condensate in COPD. COPD 2017;14:238–44.
- 59 Mathew S, Bauer KL, Fischoeder A, et al. The anergic state in sarcoidosis is associated with diminished dendritic cell function. J Immunol 2008;181:746–55.
- 60 Chopra A, Kalkanis A, Judson MA. Biomarkers in sarcoidosis. Expert Rev Clin Immunol 2016;12:1191–208.
- 61 Ascoli C, Huang Y, Schott C, et al. A circulating microRNA signature serves as a diagnostic and prognostic indicator in sarcoidosis. Am J Respir Cell Mol Biol 2018;58:40–54.
- 62 Bargagli E, Bennett D, Maggiorelli C, *et al*. Human chitotriosidase: a sensitive biomarker of sarcoidosis. *J Clin Immunol* 2013;33:264–70.
- 63 Chen ES, Song Z, Willett MH, et al. Serum amyloid A regulates granulomatous inflammation in sarcoidosis through Toll-like receptor-2. Am J Respir Crit Care Med 2010:181:360–73.
- 64 Eurelings LEM, Miedema JR, Dalm VASH, et al. Sensitivity and specificity of serum soluble interleukin-2 receptor for diagnosing sarcoidosis in a population of patients suspected of sarcoidosis. PLoS One 2019;14:e0223897.
- 65 Greene CM, Meachery G, Taggart CC, et al. Role of IL-18 in CD4+T lymphocyte activation in sarcoidosis. J Immunol 2000;165:4718–24.
- 66 Grutters JC, Fellrath J-M, Mulder L, et al. Serum soluble interleukin-2 receptor measurement in patients with sarcoidosis: a clinical evaluation. Chest 2003;124:186–95.
- 67 Kahkouee S, Samadi K, Alai A, et al. Serum ACE level in sarcoidosis patients with typical and atypical HRCT manifestation. Pol J Radiol 2016;81:458–61.
- 68 Kieszko R, Krawczyk P, Jankowska O, et al. The clinical significance of interleukin 18 assessment in sarcoidosis patients. Respir Med 2007;101:722–8.
- 69 Klech H, Kohn H, Kummer F, et al. Assessment of activity in sarcoidosis. sensitivity and specificity of 67gallium scintigraphy, serum ACE levels, chest roentgenography, and blood lymphocyte subpopulations. Chest 1982;82:732–8.
- 70 Miyoshi S, Hamada H, Kadowaki T, et al. Comparative evaluation of serum markers in pulmonary sarcoidosis. Chest 2010;137:1391–7.
- 71 Novick D, Kim S, Kaplanski G, et al. Interleukin-18, more than a Th1 cytokine. Semin Immunol 2013;25:439–48.
- 72 Ringkowski S, Thomas PS, Herbert C. Interleukin-12 family cytokines and sarcoidosis. Front Pharmacol 2014;5:233.
- 73 Su R, Li MM, Bhakta NR, et al. Longitudinal analysis of sarcoidosis blood transcriptomic signatures and disease outcomes. Eur Respir J 2014;44:985–93.

Review

- 74 Su R, Nguyen M-LT, Agarwal MR, et al. Interferon-Inducible chemokines reflect severity and progression in sarcoidosis. Respir Res 2013;14:121.
- 75 Thi Hong Nguyen C, Kambe N, Kishimoto I, et al. Serum soluble interleukin-2 receptor level is more sensitive than angiotensin-converting enzyme or lysozyme for diagnosis of sarcoidosis and may be a marker of multiple organ involvement. J Dermatol 2017;44:789–97.
- 76 Yasar Z, Özgül M, Cetinkaya E. Angiotensin-Converting enzyme as a predictor of extrathoracic involvement of sarcoidosis. Sarcoidosis Vasc Diffuse Lung Dis 2015;32:318–24.
- 77 Zhang Y, Chen X, Hu Y, et al. Preliminary characterizations of a serum biomarker for sarcoidosis by comparative proteomic approach with tandem-mass spectrometry in ethnic Han Chinese patients. Respir Res 2013:14:18.
- 78 Popević S, Šumarac Z, Jovanović D, et al. Verifying Sarcoidosis Activity: Chitotriosidase versus ACE in Sarcoidosis - a Case-control Study. J Med Biochem 2016;35:390–400.
- 79 Ungprasert P, Carmona EM, Crowson CS, et al. Diagnostic utility of angiotensin-converting enzyme in sarcoidosis: a population-based study. *Lung* 2016;194:91–5.
- 80 Du S-S, Zhao M-M, Zhang Y, et al. Screening for differentially expressed proteins relevant to the differential diagnosis of sarcoidosis and tuberculosis. PLoS One 2015;10:e0132466.
- 81 Kobak S, Semiz H, Akyildiz M, et al. Serum adipokine levels in patients with sarcoidosis. Clin Rheumatol 2020;39:2121–5.
- 82 Tanimura H, Mizuno K, Okamoto H. Serum levels of soluble CD163 as a specific marker of macrophage/monocyte activity in sarcoidosis patients. Sarcoidosis Vasc Diffuse Lung Dis 2015;32:99–105.
- 83 Kiko T, Yoshihisa A, Kanno Y, et al. A multiple biomarker approach in patients with cardiac sarcoidosis. *Int Heart J* 2018;59:996–1001.
- 84 Perez IE, Garcia MJ, Taub CC. Multimodality imaging in cardiac sarcoidosis: is there a winner? *Curr Cardiol Rev* 2016;12:3–11.
- 85 Moss BP, Patel DC, Tavee JO, et al. Evaluating S100B as a serum biomarker for central neurosarcoidosis. Respir Med 2020;162:105855.
- 86 Danila E, Jurgauskiene L, Norkuniene J, et al. BAL fluid cells in newly diagnosed pulmonary sarcoidosis with different clinical activity. Ups J Med Sci 2009;114:26–31.
- 87 Shen Y, Pang C, Wu Y, et al. Diagnostic performance of bronchoalveolar lavage fluid CD4/CD8 ratio for sarcoidosis: a meta-analysis. EBioMedicine 2016:8:302–8.
- 88 Crouser ED, Culver DA, Knox KS, et al. Gene expression profiling identifies MMP-12 and ADAMDEC1 as potential pathogenic mediators of pulmonary sarcoidosis. Am J Respir Crit Care Med 2009;179:929–38.
- 89 Mohan A, Neequaye N, Malur A, et al. Matrix metalloproteinase-12 is required for granuloma progression. Front Immunol 2020;11:553949.
- 90 Darlington P, Kullberg S, Eklund A, et al. Subpopulations of cells from bronchoalveolar lavage can predict prognosis in sarcoidosis. Eur Respir J 2020;55. doi:10.1183/13993003.01450-2019. [Epub ahead of print: 30 01 2020]
- 91 Idali F, Wikén M, Wahlström J, et al. Reduced Th1 response in the lungs of HLA-DRB1*0301 patients with pulmonary sarcoidosis. Eur Respir J 2006;27:451–9.
- 92 Lim MY, Thomas PS. Biomarkers in exhaled breath condensate and serum of chronic obstructive pulmonary disease and non-small-cell lung cancer. *Int J Chronic Dis* 2013;2013:1–15.
- 93 Leiten EO, Martinsen EMH, Bakke PS, et al. Complications and discomfort of bronchoscopy: a systematic review. *Eur Clin Respir J* 2016;3:33324.
- 94 Montuschi P. Analysis of exhaled breath condensate in respiratory medicine: methodological aspects and potential clinical applications. *Ther Adv Respir Dis* 2007;1:5–23.
- 95 Jackson AS, Sandrini A, Campbell C, et al. Comparison of biomarkers in exhaled breath condensate and bronchoalveolar lavage. Am J Respir Crit Care Med 2007;175:222–7.
- 96 Gessner C, Scheibe R, Wötzel M, et al. Exhaled breath condensate cytokine patterns in chronic obstructive pulmonary disease. Respir Med 2005;99:1229–40.
- 97 Montuschi P, Kharitonov SA, Ciabattoni G, et al. Exhaled leukotrienes and prostaglandins in COPD. *Thorax* 2003;58:585–8.
- 98 Klaassen EMM, Kant KDGvande, Jöbsis Q, et al. Symptoms, but not a biomarker response to inhaled corticosteroids, predict asthma in preschool children with recurrent wheeze. Mediators Inflamm 2012;2012:1–7.
- 99 Zietkowski Z, Tomasiak MM, Skiepko R, et al. Rantes in exhaled breath condensate of stable and unstable asthma patients. Respir Med 2008;102:1198–202.
- 100 Mohan N, Akter R, Bryant K, et al. Exhaled breath markers of alveolar macrophage activity in sarcoidosis. Inflamm Res 2016;65:471–8.

- 101 Hamsten C, Wiklundh E, Grönlund H, et al. Elevated levels of FN1 and CCL2 in bronchoalveolar lavage fluid from sarcoidosis patients. Respir Res 2016:17:69.
- 102 Ahmadzai H, Cameron B, Chui J, et al. Measurement of neopterin, TGF-β1 and ACE in the exhaled breath condensate of patients with sarcoidosis. J Breath Res 2013;7:046003.
- 103 Loke WSJ, Freeman A, Garthwaite L, et al. T-Bet and interleukin-27: possible TH1 immunomodulators of sarcoidosis. *Inflammopharmacology* 2015;23:283–90.
- 104 Alipoor SD, Mortaz E, Garssen J, et al. Exosomes and exosomal miRNA in respiratory diseases. Mediators Inflamm 2016;2016:1–11.
- 105 Kishore A, Navratilova Z, Kolek V, et al. Expression analysis of extracellular microRNA in bronchoalveolar lavage fluid from patients with pulmonary sarcoidosis. Respirology 2018;23:1166–72.
- 106 Scheideler L, Manke HG, Schwulera U, et al. Detection of nonvolatile macromolecules in breath. A possible diagnostic tool? Am Rev Respir Dis 1993;148:778–84.
- 107 Qazi KR, Torregrosa Paredes P, Dahlberg B, et al. Proinflammatory exosomes in bronchoalveolar lavage fluid of patients with sarcoidosis. *Thorax* 2010;65:1016–24.
- 108 Costabel U, Hunninghake GW. ATS/ERS/WASOG statement on sarcoidosis. sarcoidosis statement Committee. American thoracic Society. European respiratory Society. world association for sarcoidosis and other granulomatous disorders. *Eur Respir J* 1999;14:735.
- 109 Baughman RP, Nagai S, Balter M, et al. Defining the clinical outcome status (COS) in sarcoidosis: results of WASOG Task force. Sarcoidosis Vasc Diffuse Lung Dis 2011;28:56–64.
- 110 Salamo O, Mortaz E, Mirsaeidi M. Noncoding RNAs: new players in pulmonary medicine and sarcoidosis. Am J Respir Cell Mol Biol 2018;58:147–56.
- 111 Brown D, Rahman M, Nana-Sinkam SP. Micrornas in respiratory disease. A clinician's overview. *Ann Am Thorac Soc* 2014;11:1277–85.
- 112 Bak M, Jazwa A, Kasper L, et al. Involvement of microRNAs in the inflammatory pathways of pulmonary sarcoidosis. J Physiol Pharmacol 2015;66:635–42.
- 113 Bartel DP. MicroRNAs: genomics, biogenesis, mechanism, and function. Cell 2004;116:281–97.
- 114 Pattnaik B, Sryma PB, Mittal S, et al. MicroRNAs in pulmonary sarcoidosis: a systematic review. Respir Investig 2020;58:232–8.
- 115 Jazwa A, Kasper L, Bak M, et al. Differential inflammatory microRNA and cytokine expression in pulmonary sarcoidosis. Arch Immunol Ther Exp 2015;63:139–46.
- 116 Novosadová E, Chabronova A, Kolek V, et al. The Serum Expression of Selected miRNAs in Pulmonary Sarcoidosis with/without Löfgren's Syndrome. Mediators Inflamm 2016;2016:1–12.
- 117 Nana-Sinkam SP, Acunzo M, Croce CM, et al. Extracellular vesicle biology in the pathogenesis of lung disease. Am J Respir Crit Care Med 2017;196:1510–8.
- 118 Kalluri R, LeBleu VS. Discovery of double-stranded genomic DNA in circulating exosomes. Cold Spring Harb Symp Quant Biol 2016;81:275–80.
- 119 Gallo A, Tandon M, Alevizos I, et al. The majority of microRNAs detectable in serum and saliva is concentrated in exosomes. PLoS One 2012;7:e30679.
- 120 Arroyo JD, Chevillet JR, Kroh EM, et al. Argonaute2 complexes carry a population of circulating microRNAs independent of vesicles in human plasma. Proc Natl Acad Sci U S A 2011;108:5003–8.
- 121 Turchinovich A, Weiz L, Langheinz A, et al. Characterization of extracellular circulating microRNA. Nucleic Acids Res 2011;39:7223–33.
- 122 Momen-Heravi F, Saha B, Kodys K, et al. Increased number of circulating exosomes and their microRNA cargos are potential novel biomarkers in alcoholic hepatitis. J Transl Med 2015;13:261.
- 123 Kishore A, Navratilova Z, Kolek V. Detection of exosomal miRNA in pulmonary sarcoidosis. Eur Res J 2014.
- 124 Hng Y, Waters S, Chen Y. Exosome-related mediators in serum and exhaled breath condensate in sarcoidosis. Respirology 2019;24:116.
- 125 Crouser ED, Julian MW, Crawford M, et al. Differential expression of microRNA and predicted targets in pulmonary sarcoidosis. Biochem Biophys Res Commun 2012;417:886–91.
- 126 Dyskova T, Fillerova R, Novosad T, et al. Correlation network analysis reveals relationships between microRNAs, transcription factor T-bet, and deregulated Cytokine/Chemokine-Receptor network in pulmonary sarcoidosis. Mediators Inflamm 2015;2015:1–16.
- 127 Kiszałkiewicz J, Piotrowski WJ, Pastuszak-Lewandoska D, et al. Altered miRNA expression in pulmonary sarcoidosis. BMC Med Genet 2016;17:2.
- 128 Lee H, Groot M, Pinilla-Vera M, et al. Identification of miRNA-rich vesicles in bronchoalveolar lavage fluid: insights into the function and heterogeneity of extracellular vesicles. J Control Release 2019;294:43–52.

- 129 Maertzdorf J, Weiner J, Mollenkopf H-J, et al. Common patterns and diseaserelated signatures in tuberculosis and sarcoidosis. Proc Natl Acad Sci U S A 2012;109:7853–8.
- 130 Novosadova E, Chabronova A, Kolek V, et al. The serum expression of selected miRNAs in pulmonary sarcoidosis with/without Löfgren's syndrome. Mediators Inflamm 2016;2016:1–12.
- 131 Tomankova T, Zurkova M, Kolek V. Microrna profiling revealed downregulated miR-204 expression in bronchoalveolar cells from patients with pulmonary sarcoidosis. Am J Respir Crit Care Med 2020;201:A1207.
- 132 Crouser ED, Hamzeh NY, Maier LA, et al. Exosomal microRNA for detection of cardiac sarcoidosis. Am J Respir Crit Care Med 2017;196:931–4.
- 133 Landi C, Carleo A, Cillis G, et al. Sarcoidosis: proteomics and new perspectives for improving personalized medicine. Expert Rev Proteomics 2018;15:829–35.
- 134 Zhou T, Casanova N, Pouladi N, et al. Identification of JAK-STAT signaling involvement in sarcoidosis severity via a novel microRNA-regulated peripheral blood mononuclear cell gene signature. Sci Rep 2017;7:4237.
- 135 Mroz RM, Korniluk M, Stasiak-Barmuta A, et al. Increased levels of interleukin-12 and interleukin-18 in bronchoalveolar lavage fluid of patients with pulmonary sarcoidosis. J Physiol Pharmacol 2008;59 Suppl 6:507–13.
- 136 Naumnik W, Naumnik B, Niklińska W. Interleukin-33 as a New Marker of Pulmonary Sarcoidosis. In: Pokorski M, ed. *Noncommunicable diseases. Cham: Springer international publishing*, 2015: 1–6.
- 137 Thillai M, Eberhardt C, Lewin AM, et al. Sarcoidosis and tuberculosis cytokine profiles: indistinguishable in bronchoalveolar lavage but different in blood. PLoS One 2012;7:e38083.